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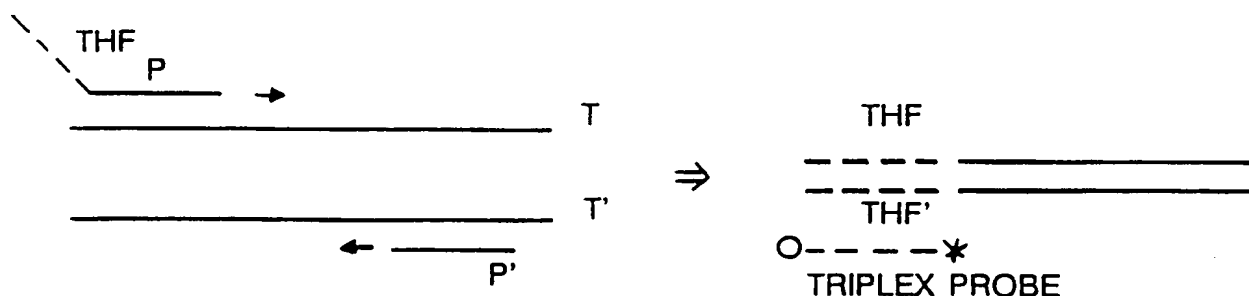
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(54) Title: NUCLEIC ACID SEQUENCE DETECTION BY TRIPLE HELIX FORMATION



(57) Abstract

Disclosed is a method for detecting a nucleic acid target sequence by formation of triple helix nucleic acid structures. The method may, but need not, involve amplifying the nucleic acid *in vitro* using cycles of denaturation and amplification to yield product duplexes, and detecting the product duplexes by hybridizing a third strand of nucleic acid to the product duplexes without denaturation. The triple helix-forming duplex sequences may be endogenous to the target sequence being detected, or they may be introduced in the probes used during amplification.

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NUCLEIC ACID SEQUENCE DETECTION BY TRIPLE HELIX FORMATIONBackground of the Invention

The invention relates to the detection of DNA sequences, particularly following amplification by the polymerase chain reaction (PCR) or other methods of selective enzymatic amplification of a target sequence.

The polymerase chain reaction (PCR) is a rapid procedure for the in vitro enzymatic amplification of a specific segment of DNA. This method has proven extremely useful in detection of infectious agents in experimental and clinical settings. PCR-amplified products may be detected by a number of methods.

Syvanen et al. (Nuc. Acid. Res. 16: 11327, 1988) describe the use of oligonucleotides, labelled with biotin at their 5' ends, as primers in a PCR reaction. The resultant amplified DNA is denatured and hybridized to a radiolabelled probe; the desired product is then captured on an avidin matrix, through an avidin-biotin interaction, and detected by the presence of the radiolabelled probe.

Kemp et al. (Proc. Natl. Acad. Sci. USA 86: 2423, 1989) describe a method for colorimetric detection of specific DNA segments amplified by PCR. One set of oligonucleotide PCR primers is covalently attached to a label (for example, biotin or a site for binding a double-stranded DNA binding protein). The amplified double-stranded DNA product can be immobilized or bound by use of a reagent which has affinity for the label (e.g., a DNA binding protein to bind the site for the DNA binding protein) and the bound DNA detected (e.g., by labelling the biotinylated DNA with avidin linked to a detectable agent,

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such as horseradish peroxidase). The presence of the detectable agent (e.g., horseradish peroxidase) on the solid phase is then assayed using a chromogenic substrate by standard techniques.

5 Keller et al. (Anal. Biochem. 177:27, 1989) describe a microtiter plate sandwich hybridization assay for detection of PCR products. A well in a microtiter plate is covalently coupled to a capture DNA. PCR-amplified target DNA is denatured, hybridized to the capture DNA, and
10 detected with a biotin-labelled detection probe.

Kenten et al. (publication of IGEN, Inc., Rockville, MD) describe use of a biotin-labelled oligonucleotide primer and an ORIGEN[®]-labelled oligonucleotide primer. Following PCR, the biotin-labelled product is caused to bind to
15 streptavidin and the presence of ORIGEN[®] assayed by electrochemiluminescent techniques.

Kumar et al. (Aids Research and Human Retroviruses 5:345, 1989) describe a probe shift assay. In this assay, PCR-amplified target DNA is denatured, hybridized to a
20 complementary radiolabelled probe, and subjected to non-denaturing polyacrylamide gel electrophoresis before and after treatment with S1 nuclease. Increased migration of the DNA band following S1 nuclease treatment indicates the presence of target DNA.

25 Summary of the Invention

In general, the invention features a method for detecting a nucleic acid. The method involves the steps of amplifying the nucleic acid in vitro using cycles of denaturation and amplification to yield product duplexes,
30 and detecting product duplexes by hybridizing a third strand of nucleic acid to the product duplexes without

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denaturation. In preferred embodiments, the amplifying may be accomplished by polymerase chain reaction or ligase chain reaction (LCR). When used with PCR amplification, the method may include amplification by use of at least two PCR
5 primers, one of the primers including a priming sequence hybridizable with the nucleic acid to be detected and one strand of a triple helix-forming sequence which need not hybridize to the original target DNA molecules, the other primer being hybridizable to the complement of the nucleic
10 acid. When used with LCR, the method may include amplification by use of two or more pairs of ligatable LCR hybridization probes; such a pair may consist of one probe which includes a sequence hybridizable to the original target molecule and a second probe which includes an
15 adjacent sequence that hybridizes to the target molecule and a triple helix-forming sequence that need not hybridize to the original target sequence but that is part of the ligated duplex molecules generated in LCR. The invention also features such PCR primers and LCR hybridization probes.

20 The invention further features a method for detecting the presence, in a biological sample, of a pathogen which includes an endogenous triple helix-forming nucleic acid sequence, involving hybridizing a third strand of nucleic acid to a duplex including the endogenous triple
25 helix-forming sequence without denaturation of the duplex. The method will work on biological samples generally prepared in accordance with known hybridization procedures.

Preferred embodiments of both methods include the following features. The third strand of nucleic acid
30 includes a polypyrimidine sequence of at least 15 nucleotides and may further include one or more purine residues, wherein each of the purine residues is flanked by 9 or more pyrimidine residues; alternatively, the third

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strand of nucleic acid includes a polypurine sequence of at least 15 nucleotides and may further include one or more pyrimidine residues, wherein each of the pyrimidine residues is flanked by 9 or more purine residues. The third strand
5 may include one or more modified residues. The third strand of nucleic acid may be covalently attached to a solid support, preferably, a microparticle. The target sequence nucleic acid is detected using an FCA format, a PCFIA format, or a microtiter well format. The third strand of
10 nucleic acid may be covalently attached to a reporter group, preferably, sulforhodamine or alkaline phosphatase. The nucleic acid may be detected by enhanced ethidium bromide fluorescence following contact with a third strand. The duplex PCR product is isolated prior to detection. More
15 than one nucleic acid duplex can be detected without separating the target duplexes, and detection of additional nucleic acid duplexes involves hybridizing each of the additional duplexes with a third strand of nucleic acid. The third strand may consist of a single molecule that
20 accomplishes specific duplex capture as well as detection. Alternatively, those two functions may be on two or more separate single-stranded molecules which hybridize to different locations on the duplex being detected and, together, make up the "third strand" as we use that term.

25 In other preferred embodiments, the nucleic acid is derived from M. paratuberculosis; the nucleic acid is derived from a retrovirus; the retrovirus is a caprine arthritis encephalitis virus; the retrovirus is a human immunodeficiency virus, and the third strand includes at
30 least 15 consecutive nucleotides of the sequence: tccccctttctttttt (SEQ ID NO.: 37) or at least 15 consecutive nucleotides of the sequence: ttaccttttccttcctttt (SEQ ID NO.: 38); the retrovirus is a feline leukemia virus, and the

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third strand includes the sequence: ttcccttttttccttt (SEQ ID NO.: 39) or includes at least 15 consecutive nucleotides of the sequence: ttttccctctggggtctcctttcccttctttct (SEQ ID NO.: 40); the retrovirus is a feline immunodeficiency virus, and the third strand includes at least 15 consecutive nucleotides of the sequence: ttcttcttcttcttttcttctt (SEQ ID NO.: 41) or at least 15 consecutive nucleotides of the sequence: ttttccttttctgttttcttcttcttcttcttctt (SEQ ID NO.: 42); the nucleic acid is derived from a human papilloma virus; the human papilloma virus is HP-16, and the third strand includes the sequence: ccccttctccc (SEQ ID NO.: 43) or at least 15 consecutive nucleotides of the sequence: tctcctcctcctactttatctacc (SEQ ID NO.: 44); the human papilloma virus is HP-18, and the third strand includes at least 15 consecutive nucleotides of the sequence: tttttatctacttccccctct (SEQ ID NO.: 45) or includes the sequence: ctccttctccttct (SEQ ID NO.: 46); and the nucleic acid is derived from a hepatitis B virus, and the third strand includes at least 15 consecutive nucleotides of the sequence: aggggaagaagaagacggcaagg (SEQ ID NO.: 47) or includes the sequence: gagggaggaaaggag (SEQ ID NO.: 48).

The invention yet further features a purified single-stranded nucleic acid probe including at least 15 consecutive nucleotides of the triple helix-forming sequence: ctctttcctctctttttcccc (SEQ ID NO.: 9) or ctctcttctctcttctctcc (SEQ ID NO.: 10).

An alternative method for detecting a nucleic acid is also featured. The method involves a competitive triple helix binding assay which includes the steps of amplifying the nucleic acid in vitro using cycles of denaturation and amplification to yield product duplexes, hybridizing a third strand of nucleic acid to the product duplexes without denaturation, hybridizing any free third strand with a

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nucleic acid hook sequence covalently bound to a solid support, and measuring the amount of product duplex as the inverse measure of the quantity of the third strand bound to the hook sequence. In preferred embodiments, the nucleic acid hook sequence is double-stranded.

The methods of the invention are incorporated into a kit for detecting a nucleic acid. The kit includes a single-stranded nucleic acid probe capable of specifically hybridizing to a triple helix-forming sequence of the nucleic acid, the probe being covalently attached to a reporter group, and a reagent for detecting the reporter group bound to the nucleic acid.

In preferred embodiments, the single-stranded nucleic acid is covalently attached to a solid support; the kit includes a second single-stranded nucleic acid capable of specifically hybridizing to a triple helix-forming sequence of the nucleic acid, and the second single-stranded nucleic acid is covalently attached to a solid support; the solid support is a microparticle; the nucleic acid is detected using a microtiter well format, an FCA format, or a PCFIA format; and the reporter group is alkaline phosphatase or sulforhodamine. To facilitate amplification, the kit may further feature at least two PCR primers, one of the primers including a priming sequence hybridizable with the nucleic acid to be detected and one strand of a triple helix-forming sequence which need not hybridize to the original target DNA molecule, the other primer being hybridizable to the complement of the nucleic acid. Alternatively, the kit may include two or more pairs of ligatable LCR hybridization probes; such a pair may consist of one probe which includes a sequence hybridizable to the original target molecule and a second probe which includes an adjacent sequence that hybridizes to the target molecule and a triple helix-forming

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sequence that need not hybridize to the original target sequence but that is part of the ligated duplex molecules generated in LCR.

By "triple helix-forming" is meant (a double-stranded nucleotide sequence) capable of specifically binding, e.g., by Hoogsteen hydrogen bonds (Hoogsteen, *Acta. Cryst.* 12:822, 1959), to a third strand of nucleic acid. Triple helix-forming sequences include, without limitation, stretches of polypurine and polypyrimidine residues. By "endogenous triple helix-forming sequence" is meant a sequence which occurs naturally in the target sequence (e.g., is not introduced by use of PCR or LCR) and is capable of forming a triple helix. By "target sequence" is meant a nucleic acid sequence, the presence or absence of which is desired to be detected. In the context of a preferred application of the methods of the invention, it is a sequence derived from a pathogenic organism. "Modified nucleotides" include, but are not limited to, protonated or methylated nucleotides, preferably, cytosine residues. By "reporter group" is meant any molecule which facilitates such target sequence detection, including, without limitation, sulforhodamine and alkaline phosphatase. By "microparticle" is meant any small solid support, including, without limitation, a Latex microparticle. By "amplification" is meant an increase in number of a particular nucleic acid sequence and may be accomplished, without limitation, by the in vitro methods of polymerase chain reaction or ligase chain reaction. By "duplex" is meant a double-stranded nucleic acid sequence. By "priming sequence" is meant a single-stranded nucleic acid which hybridizes to a single-stranded target sequence template and facilitates PCR amplification. Such a priming sequence would be included in a "PCR primer". By "LCR hybridization

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"probe" is meant one of a pair of ligatable single-stranded nucleic acid sequences which hybridizes to a single-stranded target sequence and facilitates ligase chain reaction. As used herein, the term includes a probe containing a region complementary to, and therefore capable of hybridizing to, the original target sequence (i.e., the "target-hybridizing sequence") as well as a probe which includes both a region complementary to the target sequence and a triple helix-forming sequence that need not hybridize to the original target sequence but that is part of the ligated duplex molecules generated in LCR. By "adjacent" is meant (two probes) positioned closely enough on a DNA template to be joined following a standard DNA ligation reaction. By "purified nucleic acid" is meant nucleic acid separated from other sequences with which it is naturally associated. By "probe" is meant a third strand of DNA which specifically binds, by triple helical interactions, to its complementary target sequence. By a "hook sequence" is meant a single-stranded or double-stranded oligonucleotide which hybridizes to a particular nucleic acid and facilitates its detection. The hook sequence may be covalently bound to a solid support to allow nucleic acid capture prior to detection.

The methods of the invention facilitate the rapid, specific, and automated isolation and/or detection of a nucleic acid target sequence. These methods have a number of advantages. For example, because stable triple helical complexes form at room temperature, there is no need for chemical or thermal denaturation of the double-stranded target sequence product prior to detection. In addition, isolation and/or detection of a target sequence is accomplished using triplex probes bound to capture molecules or reporter groups. Many such methods of capture and many such reporter groups exist, allowing the methods of the

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invention to be adapted to several formats, some of which are described below. Further, triplex probes hybridize efficiently only with the double-stranded target sequence. As a result, single-stranded PCR primers do not compete to any appreciable extent with target sequence for interaction with the triplex probes, and very little capture and/or detection reagent (i.e., triplex probe) is required. Finally, many different PCR products can be separately captured or detected since, in principle, any triple helix-forming sequence (for example, any polypyrimidine sequence of the appropriate length) can be used to introduce, during PCR amplification, a triple helix binding site into the target sequence. Each of these binding sites can then be separately isolated and/or detected using the methods of the invention.

Description of the Preferred Embodiments

The drawings will first briefly be described.

Drawings

Fig. 1 is a schematic representation of the PCR-directed incorporation of a triple helix-forming sequence into a target sequence and its triple helical detection.

Fig. 2 is a schematic representation of the LCR-directed incorporation of a triple helix-forming sequence into a target sequence and its triple helical detection.

Fig. 3 is a schematic representation of the IS900 target sequence and the triple helix-forming primers and flanking primers used for PCR amplification.

Fig. 4 is a graph showing triple helical detection of M. paratuberculosis-derived sequence IS900.

Fig. 5 is a graph showing detection of an endogenous triple helix-forming sequence of feline immunodeficiency virus.

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Fig. 6 is a graph showing triple helical detection of a caprine arthritis encephalitis virus-derived sequence.

Fig. 7 is a graph showing triple helical detection of a human immunodeficiency virus, type 1-derived sequence.

Fig. 8 is a list of genes whose sequences include a homopolymer tract of 25 base pairs or greater.

Methods

The methods of the invention allow detection of any target sequence containing a triple helix-forming region. Such a triple helix-forming region may be naturally-occurring (i.e., endogenous) or it may be incorporated into the target sequence during amplification *in vitro*, e.g., by polymerase chain reaction (PCR) or ligase chain reaction (LCR). PCR primers used for such an amplification reaction, termed "triplex primers" or "triple helix-forming primer", contain (1) a sequence complementary (and therefore capable of hybridizing) to single-stranded target sequence (P and P') and (2) a sequence which, when incorporated into the amplified nucleic acid, is capable of forming a triple helix (THF and THF'; Fig. 1). The basic PCR reaction is well known, and there is no need to provide a detailed description of it here (see, e.g., U.S. Patent No. 4,683,202, hereby incorporated by reference). Following amplification, the target sequence present in the resulting products is detected using a "triplex probe" (i.e., a single-stranded probe having a sequence which binds to amplified products having a duplex triple helix-forming target sequence and allows isolation of the target sequence) (Fig. 1). The triplex probes may be bound to a solid phase which facilitates capture (O, Fig. 1). In one example, this solid phase is a Latex microparticle which may be isolated by centrifugation or by capture on a fluorescence concentration analyzer (FCA) plate following vacuum

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filtration (see below). In addition, the triplex probe may include a "detection sequence" (i.e., a sequence which binds to a duplex target sequence and allows the target sequence to be assayed). The detection sequence is generally bound to a reporter group (*, Fig. 1), which indicates the presence of the duplex target sequence. In one example, this reporter group is sulforhodamine, a substrate which is detected by fluorescence assay. A single triplex probe can be bound both to a solid phase and to a reporter group. Alternatively, two triplex probes can be used, one bound to a solid phase and a second bound to a reporter group. In the latter case the solid phase-bound probe and the reporter-bound probe bind at different triple helix-forming sites on the duplexes to be detected.

The LCR reaction is similarly well known (see, e.g., European Patent Application No. 0 336 731 A2, hereby incorporated by reference). LCR incorporation of a triple helix-forming region is accomplished in two steps. First, target sequence DNA is denatured and to the appropriate single strand is annealed a pair of LCR hybridization probes (e.g., LCR probe 1 and LCR probe 2) which are complementary to adjacent sequences (a and b, Fig. 2). In one example, LCR probe 1 includes a "target-hybridizing sequence" (P_a , Fig. 2) and one strand of a triple helix-forming sequence (THF, Fig. 2). LCR probe 2 includes an adjacent target-hybridizing sequence (P_b , Fig. 2) and a detectable label (e.g., sulforhodamine; *, Fig. 2). In the same annealing reaction, another pair of adjacent probes are hybridized to the opposite strand of target DNA. In this case, LCR probe 1' includes a target-hybridizing sequence (P_a' , Fig. 2) and one strand of the triple helix-forming sequence (THF', Fig. 2); and LCR probe 2' includes a target-hybridizing sequence

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(P_b', Fig. 2), and it may be bound to the same detectable label (*; Fig. 2). LCR probes 1 and 2 and LCR probes 1' and 2' are ligated together, and the duplexes are denatured and re-annealed, allowing complex formation between the newly-ligated probe strands. Capture is accomplished with a third strand of nucleic acid (i.e., a "triplex probe", Fig. 2) which forms a triple helix with the double-stranded triple helix-forming sequence. The third strand may, e.g., be attached to a solid support (O, Fig. 2), such as a microparticle, to aid in product duplex isolation (e.g., as described herein). Triple helices are assayed, e.g., by measuring sulforhodamine fluorescence.

In general, the duplex-binding sequences of the triplex probes used in these studies are stretches of polypyrimidine or polypurine residues of identical polarity and complementary to the polypurine or polypyrimidine tract (respectively) of the target sequence to be detected. These polypurine or polypyrimidine sequences are, preferably, 15 nucleotides or greater in length, optimally, 20 nucleotides. Such probe sequences cannot form a stable Watson-Crick complex or parallel helical complex (i.e., Hoogsteen complex) with the single-stranded primers but can stably interact only with a duplex primer sequence bound to its Watson-Crick partner (i.e., incorporated into the sequence of the double-stranded target sequence). For example:

→ 5' ggaaggaaagaaggag 3' Target sequence (SEQ ID NO.: 1)
- 3' CCTTCCTTTCTTCCTC 5' Triplex Primer (SEQ ID NO.: 2)
5' CCTTCCTTTCTTCCTC 3' Triple Helical Probe
(SEQ ID NO.:3)

This observation underlies the utility of the invention; a third strand probe does not efficiently

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interact with a single-stranded primer sequence until such a sequence is incorporated into a double-stranded PCR product.

Reaction conditions for triple helix formation have been determined and are described herein and in, for example, Griffin and Dervan (*Science* 245:967, 1989); Moser and Dervan (*Science* 238:645, 1987); Lyamichev et al. (*Nucl. Acids Res.* 16:2165, 1988); Strobel et al. (*J. Am. Chem. Soc.* 110: 7927, 1988); Povsic and Dervan (*J. Am. Chem. Soc.* 111:3059, 1989); and Maher et al. (*Science* 245:725, 1989).

Triple helix-forming sequences are not limited to polypyrimidine or polypurine stretches. Certain purine-pyrimidine-purine tracts also form stable triple helices and provide additional sequence versatility for this approach. For example, an adenosine residue interrupting a polypyrimidine target sequence can be accommodated with a guanosine residue in the triplex probe strand (Griffin and Dervan, *Science* 245:967, 1989). Base pair mismatches may be compensated for by adjustment of solution conditions, for example, by maintaining a pH of between 6.2 and 7.0, and/or by inclusion of 10% ethanol, and/or by adjustment of the reaction temperature to 23°C or below (for example, 0°C) (see, for example, Griffin and Dervan, *Science* 245:967, 1989). Triple helical interactions are also improved by binding an intercalating drug such as acridine to the end of the single-stranded triplex probe strand (i.e., the third strand of nucleic acid). Upon binding to a duplex target sequence, the triplex probe-bound drug intercalates into the duplex DNA and contributes to the total binding constant. There are reports that the use of a third strand of nucleic acid bound to an intercalating drug allows specific triple helical interactions using third strands of DNA which are of a shorter length, e.g., 8 to 10 nucleotides of polypyrimidine. In addition, triple helix-forming sequences

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or triplex probe sequences may include one or more modified nucleotide bases, for example, protonated, methylated, or halogenated bases, e.g., cytosines.

Analysis of any number of DNA target sequences in a single PCR reaction mixture, or amplificate, may be accomplished using the methods of the invention. By incorporating different capture sequences into the PCR primers, a single PCR reaction may be run and target sequences may be isolated separately and detected using one or more reporter group-labelled detection sequences. Alternatively, by incorporating a common capture sequence into the PCR primers, many target sequences may be isolated simultaneously and then individually assayed using unique detection sequences, each bound to a different reporter group. Such reporter groups would be differentiated, for example, by measuring unique fluorescence properties (i.e., each reporter group is detected at a different wavelength).

The methods of the invention allow detection of any target sequence (i.e., from any organism) whose sequence, or a portion thereof, has been determined. PCR primers and triplex probes would be designed as described herein and amplification and detection of the sequence, similarly, carried out by the methods herein. The methods of the invention are particularly well-suited to the detection of retroviruses, for example, lentiviruses. Inspection of many retroviral sequences reveals the presence of unique triple helix-forming sequences, often in the viral LTR. For example, the following viral genomes include one or more triple helix-forming sequences: feline immunodeficiency virus, feline leukemia virus; human immunodeficiency virus, type 1; hepatitis B virus; and human papilloma viruses, HP-16 and HP-18.

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A very small quantity of target sequence (e.g., nucleic acid isolated from a pathogenic organism) is required for detection by these methods. The nucleic acid to be amplified and/or detected may be isolated from tissue, blood, fecal samples or the like by techniques well known to those skilled in the art.

A kit which facilitates triple helical detection of target sequences may encompass any or all of the embodiments described herein.

Experimental Information

The present invention will be further illustrated by the following examples. In the first set of examples, a triple helix-forming sequence is introduced by PCR into M. paratuberculosis-derived nucleic acid. The sequence is then detected using either a microtiter well or a fluorescence concentration assay (FCA) plate format and the reporter groups, alkaline phosphatase or sulforhodamine (Texas Red). In a separate set of examples, an endogenous triple-helix forming sequence is exploited for detection of feline immunodeficiency virus (FIV). Finally, to illustrate the general utility of the method, exogenous triple helix-forming sequences (incorporated by PCR) are exploited for detection of caprine encephalitis arthritis virus (CEAV) and human immunodeficiency virus, type 1 (HIV-1), using an FCA plate format and a sulforhodamine reporter group. These examples are not limiting to the invention.

Triple helical detection of PCR-amplified nucleic acid target sequences

PCR primers specific for the M. paratuberculosis (M.pt. or M.pt.)-derived sequence IS900 were prepared; the primer sequences were based on the published M.pt. sequence described in Green et al. (Nucl. Acids Res. 17:9063, 1989)

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and were of the following sequence (5' - 3'). (The first nucleotide of the M. pt.-specific sequence is underlined):

cccctttttctctcctttctcggacaatgacggttacgg Triplex IS210A
(SEQ ID NO.: 4)

cctctctttctctcttctctcggacaatgacggttacgg Triplex IS210B
(SEQ ID NO.: 5)

cccctttttctctcctttctcgaaggcgatcagcaacgcgg Triplex IS435A
(SEQ ID NO.: 6)

cggacaatgacggttacgg M.pt.X
(SEQ ID NO.: 7)

caaggcgatcagcaacgcgg M.pt.Y
(SEQ ID NO.: 8)

ctctttcctctctttttccccc-NH₂ Triplex Probe A
(SEQ ID NO.: 9)

ctctcttctctcttctctcc-NH₂ Triplex Probe B
(SEQ ID NO.: 10)

The triplex primers contain two domains: (1) a sequence at the 3' end specific for PCR priming of the complementary sequences in the M.pt. genomic DNA and (2) a polypyrimidine sequence at the 5' end which provides a triple helix-forming sequence. Triplex probe A (SEQ ID NO.: 9) is bound to a solid support and constitutes a capture sequence; triplex probe B (SEQ ID NO.: 10) is bound to a reporter group and constitutes a detection sequence. In other cases, the target sequence-specific region and the triple helix-forming sequence may overlap in part or in full (for example, in the case of lentiviral endogenous polypurine sequences, see below).

Oligonucleotide primers were synthesized using standard phosphoramidite chemistry on an Applied Biosystems 381-A DNA synthesizer. All nucleic acids were stripped from their controlled pore glass supports and deblocked using

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concentrated reagent ammonium hydroxide and an 8-16 hr. standard incubation time at 55°C. Oligonucleotides were desalted by gel exclusion chromatography and transferred to 0.01M MES buffer, pH 5.5 using Sephadex G10 column chromatography. Amino groups were added to the 3' termini of the primers using a 3' amino-on controlled pore glass support (Clontech, Inc.; Palo Alto, CA).

M.pt. genomic DNA was prepared as follows. M.pt. cells were disrupted by heating at 120°C for 5 min. in 0.2N NaOH, and DNA was isolated from the cell debris by standard phenol/chloroform extraction. PCR amplification of M. pt. DNA were performed using one or two of the triple helix-forming primers, triplex IS210A (SEQ ID NO.: 4), triplex IS210B (SEQ ID NO.: 5), or triplex IS435A (SEQ ID NO.: 6), above. In addition, control PCR product was prepared using primers, M.pt.X (SEQ ID NO.: 7) and M.pt.Y (SEQ ID NO.: 8), which included IS900-derived sequences but lacked any triple helix-forming sequence. PCR reactions were conducted using the standard reaction conditions recommended by Cetus (Perkin-Elmer Cetus, Norwalk, CT) and included 0.7 μ m primer. Primer concentrations were estimated by determination of absorption at 260 nm. Individual primer millimolar extinction values were calculated using the high temperature molar extinction coefficient for homopolymer strands.

Probe sequences were bound to Latex microparticles (also referred to as Latex beads) as follows. Latex microparticles (i.e., 1.0 μ carboxylate microspheres; Polysciences, Inc., Warrington, PA) bearing carboxyl groups were activated in situ by incubating for 2 hr. at pH 5.5 with a 5:1 molar ratio (with respect to carboxyl groups) of 1-ethyl-3-(3-dimethylaminopropyl)-carbodiimide (EDAC; Sigma Chemical Co., St. Louis, MO). Probe sequences bearing a

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single primary amino group (incorporated during automated chemical synthesis) were mixed with the activated Latex microparticles and the reaction was allowed to proceed for 15 min. at 25°C in 0.1M MES, pH 5.5. The reaction was terminated by the addition of excess 0.1M glutamic acid (i.e., 100 µl glutamic acid/ 1 ml reaction mixture).

Microparticles were washed 4 times with excess 0.1M MES, pH 5.5 and one time with water. The beads were resuspended in water to a density of 2.5% (w/v) solids based on original amounts used.

Probe sequences may be bound to alkaline phosphatase (AP) by the method of Jablonski et al. (*Nucl. Acids Res.* 14:6115, 1986)

EXAMPLE 1

PCR-amplified M.pt.-derived sequence IS900 was detected using Probe A (SEQ ID NO.: 9)- or Probe B (SEQ ID NO.: 10)-labelled Latex microparticles and a polyacrylamide gel format as follows. M.pt. genomic DNA was amplified using either triplex IS435A (SEQ ID NO.: 6) and triplex IS210A (SEQ ID NO.: 4) to produce PCR A=A, or triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5) to produce PCR A=B. Following PCR amplification, 5 µl of the amplificate was mixed with 10 µl of 0.5µ Latex microparticles and 4 µl 5X AP triple helix buffer (1.6% Tween 20; 10mM spermine; 1.0M MES, pH5.5) and 1 µl H₂O. The reaction was carried out at room temperature for 30 min. Triple helix formation was assessed following electrophoretic separation of bound (i.e., triple helical) product from free product on a 4% non-denaturing polyacrylamide gel. PCR products were visualized with ethidium bromide.

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Table 1 shows that triplex probes A (SEQ ID NO.: 9) and B (SEQ ID NO.: 10) specifically recognize and bind the appropriate products present in a PCR product mixture.

TABLE 1

Probe A ^a -microparticles	-	+	-	-	+	-
Probe B ^b -microparticles	+	-	-	+	-	-
PCR A=A ^c	+	+	+	-	-	-
PCR A=B ^d	-	-	-	+	+	+
Expected Binding	-	+	-	+	+	-
Observed Binding	-	+	-	+	+	-

^aSEQ ID NO.: 9

^bSEQ ID NO.: 10

^cPCR amplificate prepared as described above using M.pt. genomic DNA and primer, triplex IS435A (SEQ ID NO.: 6) and triplex IS210A (SEQ ID NO.: 4).

^dPCR amplificate prepared as described above using M.pt. genomic DNA and primers, triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5).

EXAMPLE 2

Triple helical interactions were also assessed by a sandwich assay using labelled carboxy-Latex microspheres (i.e., Latex microparticles) in a microtiter well format; triple helices were visualized using alkaline phosphatase-labelled triplex probes. M.pt. genomic DNA was amplified using either triplex primers IS435A (SEQ ID NO.: 6) and IS210B (SEQ ID NO.: 5) (to yield the triple helix-forming product, PCR A=B) or standard primers M.pt.X (SEQ ID NO.: 7)

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and M.pt.Y (SEQ ID NO.: 8) (to yield the product, M.pt. X=Y). 5 μ l of PCR amplificate was mixed with 10 μ l of 0.5 μ Probe A (SEQ ID NO.: 9)-labelled Latex microparticles, 1 μ l of varying amounts of Probe B (SEQ ID NO.: 10)-labelled alkaline phosphatase, 4 μ l 5X AP triple helix buffer, and 5 μ l H₂O. Triple helix formation was allowed to proceed for 30 minutes at room temperature. Following incubation, the reaction was centrifuged for 2 min. at 14,000 rpm in a microcentrifuge, and the supernatant was removed. Microparticles were then washed twice with 1X Enzyme Amplified Capture Buffer (EAC Buffer; 1M MES, pH 5.5, 0.02% w/v sodium azide, 1M NaCl, 0.01 Spermine, 1.5% w/v Tween 20), followed by microparticle isolation by centrifugation as above. Microparticles were then resuspended in 50 μ l Diethanolamine Enzyme Assay Buffer (DEA Buffer; 0.1M diethanolamine, 5mM MgCl₂, pH 9.5) and transferred to 96-well microtiter plates (Dynatech Inc., Chantilly, VA). Colorimetric enzyme assay reaction was initiated by the addition of 50 μ l Alkaline Phosphatase Assay Buffer (AP Buffer; 0.1M diethanolamine, pH 9.5, 5mM MgCl₂, 6 mg/ml paranitrophenol phosphate) at room temperature. Alkaline phosphatase activity was assayed over a 20 min. interval in a Molecular Devices V_{max} microtiter plate reader (Molecular Devices Inc., Menlo Park, CA). Table 2 shows that the PCR-amplified product, PCR A=B, which contains triple helix-forming sequences is detected using triplex probes A (SEQ ID NO.: 9) and B (SEQ ID NO.: 10). The control product, M.pt. X=Y, which does not contain a triple helix-forming sequence is not detected under these assay conditions.

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TABLE 2

Probe B-Alkaline phosphatase ^a (dilution)	1/5	1/10	1/50	1/5	1/10	1/50
PCR A=B ^b	5	5	5			
<u>M.pt.</u> X=Y ^c				5	5	5
Expected Color	+	+	+	-	-	-
Observed Color	+	+	+/-	+/-	-	-

^aDilution of a 1 µg/ml Probe B (SEQ ID NO.: 10) sequence bound to alkaline phosphatase.

^bPCR amplificate prepared as described above using M.pt. genomic DNA and primers, triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5).

^cPCR amplificate prepared as described above using M.pt. genomic DNA and primers, M.pt.X (SEQ ID NO.: 7) and M.pt.Y (SEQ ID NO.: 8).

To determine the specificity of the triple helical interactions, PCR amplicates described above were assayed using either Probe A (SEQ ID NO.: 9)- or oligo-dT-labelled microparticles. Binding was detected (as above) by measuring alkaline phosphatase activity. A (+) value was defined as visible color; a (-) value was defined as a white background.

Table 3 shows that Probe A (SEQ ID NO.: 9), but not oligo (dT), selectively detected PCR products amplified

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using primer triplex IS210A (SEQ ID NO.: 4) and/or triplex IS435A (SEQ ID NO.: 6).

TABLE 3

Probe A ^a -microparticles	+	+	+	-	-	-
Oligo dT-microparticles	-	-	-	+	+	+
Probe B ^b -alkaline phosphatase	+	+	+	+	+	+
PCR A=B ^c	+			+		
PCR A=A ^d		+			+	
<u>M.pt.</u> X=Y ^e			+			+
Expected Color	+	-	-	-	-	-
Observed Color	++	+	+/-	+/-	+/-	-

^aSEQ ID NO.: 9

^bSEQ ID NO.: 10

^cPCR amplificate prepared as described above using M.pt. genomic DNA and primers, triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5).

^dPCR amplificate prepared as described above using M.pt. genomic DNA and primer, triplex IS210A (SEQ ID NO.: 4) and triplex IS435A (SEQ ID NO.: 6).

^ePCR amplificate prepared as described above using M.pt. genomic DNA and primers, M.pt.X (SEQ ID NO.: 7) and M.pt.Y (SEQ ID NO.: 8).

PCR product capture and detection was dependent upon the quantity of Probe A (SEQ ID NO.: 9)-labelled beads added to the reaction mixture. This result is shown in Table 4. A 30 μ l reaction contained 10 μ l PCR amplificate, 1 μ l of a

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1:10 dilution of Probe B (SEQ ID NO.: 10)-labelled alkaline phosphatase (of concentration 0.1 $\mu\text{g/ml}$) and decreasing amounts (ranging from 0.25 to 0.10 $\mu\text{g/ml}$) of Probe A (SEQ ID NO.: 9)-labelled 1 μ Latex microparticles in 17 μl of water and 6 μl of 5X AP triple helix buffer. Triple helix formation was allowed to proceed for 30 minutes at room temperature. Following incubation, the reaction was centrifuged for 1 minute at 14,000 rpm in a microcentrifuge, the supernatant was removed, and the microparticles were washed twice with 1X Enzyme Amplified Capture Buffer (as described above) followed by particle isolation by centrifugation (at 14,000 rpm for 2 min.). The microparticles were then resuspended in 50 μl Diethanolamine Enzyme Assay Buffer and transferred to microtiter wells (as described above). The colorimetric enzyme assay reaction was initiated by the addition of 50 μl Alkaline Phosphatase Assay Buffer. Alkaline phosphatase was assayed over a 20 minute interval in a Molecular Devices V_{max} microtiter plate reader. Values shown in Table 4 were calculated from initial velocity measurements. The numbers represent MOD/min. Table 4 shows that the PCR product resulting from amplification using triplex primers IS435A (SEQ ID NO.: 6) and IS210B (SEQ ID NO.: 5) (i.e., PCR A=B) was detected with varying amounts of probe A (SEQ ID NO.: 9)-labelled Latex microparticles.

TABLE 4

Volume of Probe A ^a -microparticles	5 μl	4 μl	3 μl	2 μl
PCR A=B ^b	139.7*	113.4	156.7	102.7
M.pt. X=Y ^c	22.3	4.4	5.3	4.0

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^aSEQ ID NO.: 9

^bPCR amplificate prepared as described above using M.pt. genomic DNA and primers, triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5).

^cPCR amplificate prepared as described above using M.pt. genomic DNA and primers, M.pt.X (SEQ ID NO.: 7) and M.pt.Y (SEQ ID NO.: 8).

To assess the "worst case" contribution of background binding (i.e., by primer dimer accumulation in the absence of target sequence nucleic acid), M.pt. genomic DNA target sequence was PCR-amplified using primers, triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5) (+). A negative amplificate (-) was prepared by amplifying the primers, triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5) in the absence of any added target sequence. (+) and (-) PCR amplificates were mixed in varying proportions to a constant volume of 5 μ l. Detection was performed in a 25 μ l reaction mixture containing the mixed amplificates, 4 μ l Probe A (SEQ ID NO.: 9)-labelled microparticles, 1 μ l of a 1:10 dilution of Probe B (SEQ ID NO.: 10)-labelled alkaline phosphatase, and 5 μ l of 5X AP triple helix buffer. Reaction mixtures were processed as described above, and alkaline phosphatase activity was measured in mOD/min.

TABLE 5

+ PCR A=B(μ l)	0	1	2	3	4	5
- PCR A=B(μ l)	5	4	3	2	1	0
Alkaline phosphatase activity	32.0	43.7	34.5	43.1	79.2	85.1

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Table 5 shows that in the "worst case" (i.e., at 5 μ l PCR A=B: 5 μ l negative control), the signal/background ratio was about 2.7:1.

EXAMPLE 3

Triple helical formation was also detected using an Fluorescence Concentration Analyzer (FCA) format. The assay was performed as described for Table 2. Following triple helix formation, the reaction mixture was transferred to an FCA plate (Pandex Corp., Mundelein, IL) held under vacuum. The microparticles were then washed once with 50 μ l 2x AP triple helix buffer. 30 μ l of DEA Buffer was added, and the plate was photographed following a 60 minute incubation. Results of this analysis are shown in Table 6. A (+) value was defined as visible color; a (-) value was defined as a white background.

TABLE 6

Probe A ^a -microparticles		+	+	+	+	+	+
Probe B ^b -alkaline phosphatase	+	+	+	+	+	+	
PCR A=B ^c	+	+	+	-	-	-	
<u>M.pt.</u> X=Y ^d	-	-	-	+	+	+	
Expected Color	+	+	+	-	-	-	
Observed Color	+	+	+	-	-	-	

^aSEQ ID NO.: 9

^bSEQ ID NO.: 10

^cPCR amplicate prepared as described above using M.pt. genomic DNA and primers, triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5).

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^dPCR amplificate prepared as described above using M.pt. genomic DNA and primers, M.pt.X (SEQ ID NO.: 7) and M.pt.Y (SEQ ID NO.: 8).

EXAMPLE 4

Sulforhodamine (Texas Red) may also be used as a sensitive reporter group to detect triple helices. In this example, PCR-amplified products were captured using Latex microparticles labelled with Triplex Probe A (SEQ ID NO.: 9) and detected with Sulforhodamine-labelled derivatives of Triplex Probe B (SEQ ID NO.: 10). Probe DNA was bound to sulforhodamine as follows. Probe DNA was transferred to 0.1M Borate buffer, pH 9.7, by PD-10 gel exclusion chromatography. 100 nmoles of oligonucleotide was reacted with 1 mg. of sulforhodamine (Molecular Probes Inc., Eugene, OR) for 2 hrs. at 4°C. The labelled oligonucleotide was purified using a PD-10 column previously blocked with a 2% solution of bovine serum albumin (Sigma Co., St. Louis, MO) in 0.1M MES buffer, followed equilibration with 0.1M MES buffer, pH 5.5.

M.pt. genomic DNA was PCR amplified using primers, triplex IS435A (SEQ ID NO.: 6) and triplex IS210B (SEQ ID NO.: 5) (producing PCR A=B). A 300 µl reaction mixture containing 30 µl of PCR amplificate, 30 µl of 5X FCA triple helix buffer (5X FCA Buffer; 1M MES, pH 5.5, 0.2% w/w sodium azide, 0.01M Spermine, 8% v/v Tween 20), 80 pmol of Sulforhodamine-labelled Probe B (SEQ ID NO.: 10), and 0.03% (w/v) Probe A (SEQ ID NO.: 9)-labelled Latex microparticles was incubated at room temperature. Aliquots were taken at the indicated time points and adsorbed onto an FCA plate held under vacuum. Following a 30 second vacuum step, the plates were washed once with 1x triple helix buffer, dried, and read in a fluorescence concentration analyzer (PANDEX

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Corp., Mundelein, IL) at an excitation wavelength of 590nm and an emission wavelength of 620nm. Results are shown in Table 7; values are expressed in relative fluorescence units.

TABLE 7

<u>Time of Reaction (min)</u>	0.5	1.0	5.0	15.0
Fluorescence	446	792	926	948

To test the specificity of triple helix formation, a 20 μ l analytical mix containing 4 μ l of 5X FCA Buffer was added to varying amounts of either a PCR A=B or M.pt. X=Y PCR amplificate. Samples were processed for FCA analysis as described above for Table 7. Results are shown in Table 8. The bottom line shows that the "worst case" signal to background ratio is 2.7:1.

TABLE 8

PCR A=B (μ l)	0 μ l	1 μ l	3 μ l	5 μ l
Fluorescence	460	1916	5416	7444
<u>M.pt.</u> X=Y (μ l)	0 μ l	1 μ l	3 μ l	5 μ l
Fluorescence	-	756	1762	2778
Signal/Background	-	2.5	3.1	2.7

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EXAMPLE 5

The primary source of background signal and, as such, the component limiting detection sensitivity is the production of "primer dimers". In the examples above, such dimers form between triplex IS210B (SEQ ID NO.: 5) and triplex IS435A (SEQ ID NO.: 6). To increase detection sensitivity, target DNA was subjected to two rounds of PCR amplification: a first round using standard flanking primers and a second round using nested triple helix-forming primers as follows. "Triple helix-forming" primers (i.e., 3 and 4 of Fig. 3) were designed to amplify the region encompassed by nucleotides 843 to 1004. "Flanking primers" (i.e., 1 and 2 of Fig. 3) were designed to amplify sequences outside of and including the triple helix-forming primers, i.e., the region encompassed by nucleotides 854 to 106. In general, these primers may be of any sequence complementary to the genomic sequence and located outside the region amplified by the triple-helix forming primers.

Samples of IS900 sequence contained in the plasmid PMB22 were diluted in a series of six 10-fold steps. Flanking primers 1 (SEQ ID NO.: 11) and 2 (SEQ ID NO. 12), of sequence shown below, were used to amplify the IS900 sequence between them; this sequence included the binding sites for the triple helix-forming primers 3 (SEQ ID NO.: 13) and 4 (SEQ ID NO.: 14) of sequence shown below.

5' gcccgcaacgccgatacc 3' IS900 Flanking primer 1
(SEQ ID NO.: 11)

5' cccaggatgacgccgaat 3' IS900 Flanking primer 2
(SEQ ID NO.: 12)

5' cccctttttctctccttttctcgccg
ctaacgcccaacac 3' IS900 Nested triple helix-
forming primer 3

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5' cctctcttctctcttctctcgct
cctcgatcatcgc 3'

(SEQ ID NO.: 13)

IS900 Nested triplex helix-
forming primer 4

(SEQ ID NO.: 14)

Following 35 cycles of flanking primer (SEQ ID NO. 11 and 12) amplification (carried out as described above), 10 μ l of the reaction mixture was amplified in a 90 μ l PCR mix using the triple helix-forming primers 3 (SEQ ID NO.: 13) and 4 (SEQ ID NO.: 14) and another 15 cycles of amplification (1 cycle is 94°C for 1 min, followed by 45° for 45 min, followed by 72°C for 45 min). 6 μ l of the final PCR amplificate was then mixed with 24 μ l of standard Analytical Mix (2.4 μ l of 2.5% Triplex probe A (SEQ ID NO. 9)-labelled Latex microparticles, i.e., approximately 100 pmoles Triplex probe A (SEQ ID NO.: 9) strand; 2.4 μ l Sulforhodamine labelled-Triplex probe B (SEQ ID NO.: 10) of concentration 0.8-1.0 pmol/ μ l; 4.8 μ l 5X Triple Helix Formation Buffer, i.e., 1M MES, 8% (v/v) Tween 20, 10mM Spermine, 0.01% (w/v) sodium azide; 1.6 μ l of 0.0025% (w/v) NYO fluorescent microspheres; and 12.8 μ l H₂O). The mixture was incubated at room temperature for 30 min and then transferred to an FCA plate, and the results were read using an excitation wavelength of 590nm and emission wavelength of 620nm.

Figure 4 shows that IS900 was selectively detected by FCA of the PCR amplificate generated with nested triple helix-forming primers 3 (SEQ ID NO. 13) and 4 (SEQ ID NO.: 14) and flanking primers, 1 (SEQ ID NO.: 11) and 2 (SEQ ID NO.: 12) (Δ). No significant detection of the IS900 sequence was observed in the PCR amplificate generated with flanking primers 1 (SEQ ID NO.: 11) and 2 (SEQ ID NO.: 12) alone (Δ).

EXAMPLE 6

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In many cases, organisms contain endogenous sequences capable of forming triple helices with a single-stranded probe sequence. Feline immunodeficiency virus (FIV) contains such sequences (Talbot et al., *Proc. Natl. Acad. Sci USA* 86:5743, 1989). These sequences were used to design primers for PCR amplification and triplex probes for detection as follows.

Feline genomic DNA containing proviral copies of feline immunodeficiency virus was prepared from FIV-infected CRFK cells by the method of Blin and Stafford (*Nucl. Acids Res.* 3:2303, 1976). Triplex Primer A (SEQ ID NO.: 15), complementary to nucleotides 9003 to 9020, and Primer 2 (SEQ ID NO.: 16), complementary to nucleotides 8918 to 8935 were designed (as described above) using the published feline immunodeficiency virus sequence of Talbot et al. (above).

5' cgaatcaaatacaactaa cccctttttctctcctttc 3'	<u>Triplex Primer A</u> (SEQ ID NO.: 15)
5' tctaactctgtcatcatc 3'	<u>Primer 2</u> (SEQ ID NO.: 16)
5' ttcttcttcttcttcttctt-NH ₂ 3'	<u>Endogenous Triplex Probe P</u> (SEQ ID NO.: 17)

Feline genomic DNA of concentration 0.5 µg/µl and including 5 x 10⁵ copies of FIV provirus was subjected to PCR amplification (by the method described in Example 5) using Triplex Primer A (SEQ ID NO.: 15) and Primer 2 (SEQ ID NO.: 16). 6 µl of the resulting amplificate was mixed with 24 µl of Standard Analytical Mix containing Triplex Primer A (SEQ ID No.: 15)-labelled Latex microparticles and sulforhodamine labelled-Triplex probe P (SEQ ID NO.: 17). FIV sequences were detected in PCR amplificates which were

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generated using either 0.4 μ M (\square , Fig. 5) or 0.8 μ M (+, Fig. 5) primer concentration. Results of these analyses are shown in Fig. 5. The endogenous FIV triple helix-forming sequence was selectively detected by Triplex Probe P (SEQ ID No.: 17) in this assay.

FIV contains a second putative triple helix-forming sequence (i.e., aaaaggaaaagacaaagaagaagaagaagaagaaa; nucleotides 8956-8992; SEQ ID NO.: 49). This sequence may be used to design PCR primers and a complementary triplex probe for detection of an FIV target sequence as described above.

EXAMPLE 7

To illustrate the generality of the methods of the invention, triple helix-forming primers and flanking primers were developed for detection of caprine encephalitis arthritis virus (CEAV) and human immunodeficiency virus 1 (HIV-1). Viral polymerase (pol) genes sequences, obtained from the Genbank database, were used to design the following primers for PCR amplification:

CEAV:

5' taggaaaggcacccccacattgg 3'	<u>CAEV Flanking primer 1</u> (SEQ ID NO.: 18)
5' cccctaagatctcctccatgg 3'	<u>CAEV Flanking primer 2</u> (SEQ ID NO.: 19)
5' cccctttttctctccttttctcat taggacttccgcatccgg 3'	<u>CAEV Nested triple helix-forming primer 3</u> (SEQ ID NO.: 20)
5' cctctcttctctcttctctcgca agtgtactctcgatatgg 3'	<u>CAEV Nested triple helix forming primer 4</u> (SEQ ID NO.: 21)

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HIV-1:

5' gcacttttaaattttcccattagtc 3'

HIV Flanking primer 1
(SEQ ID NO.: 22)

5' cctgcgggatgtggtattcc 3'

HIV Flanking primer 2
(SEQ ID NO.: 23)5' cccctttttctctcctttctcaa
gccaggaatggatggcc 3'HIV Nested triple helix-
forming primer 3
(SEQ ID NO.: 24)5' cctctcttctctcttctctccag
aagtcttgagttctcc 3'HIV Nested triple helix-
forming primer 4
(SEQ ID NO.: 25)

CAEV DNA was obtained as a plasmid clone from the LCMB recombinant clone collection (DNA #3041N); HIV-1 DNA was obtained from the American Type Culture Collection (Bethesda, MD). DNA samples were subjected to PCR amplification (by the method described in Example 5), and the resulting amplificate probed with a third strand of DNA using a Standard Analytical Mix, also as described in Example 5. For both CAEV and HIV-1 assays, Probe A (SEQ ID NO.: 9)-labelled Latex microparticles and sulforhodamine labelled-Triplex probe B (SEQ ID NO.: 10) were used for detection. Results of these analyses are shown in Fig. 6 (CAEV) and Fig. 7 (HIV-1). CAEV and HIV-1 target sequences were detected in each case by this assay.

EXAMPLE 8

The procedure described in Example 6 could also be used to test for the presence the organisms: human immunodeficiency virus, type 1 (HIV-1), feline leukemia virus (FELV), human papilloma virus-16 (HPV-16), human papilloma virus-18 (HPV-18), and hepatitis B virus (HBV). The genomic sequence of each of these organisms includes an

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endogenous sequence(s) predicted to form a triple helical structure. Such sequences include the following (5' to 3'):

HIV-1:

aggggggaaagaaaaaa (nucleotides 404-419; SEQ ID NO.: 26);

aaatggaaaaggaaggga (nucleotides 2250-2270; SEQ ID NO.: 27);

FELV:

aagggaagaaaggaaa (nucleotides 1927-1941; SEQ ID NO.: 28);

aaaaggagaccccagaggaaagggaagaaaga
(nucleotides 2147-2179; SEQ ID NO.: 29);

HPV-16:

agaggaggaggatgaaatagatgg (nucleotides 657-680; SEQ ID NO.: 30);

ggggaagagg (nucleotides 889-899; SEQ ID NO.: 31);

HPV-18:

aaaaatagatgaagggggaga (nucleotides 2227-2247; SEQ ID NO.: 32);

gaggaagaggaaga (nucleotides 2804-2817; SEQ ID NO.: 33);

HBV:

tccccttcttcttctgccgttcc (nucleotides 1490-1512; SEQ ID NO.: 34);

ctccctcctttcctc (nucleotides 2508-2522; SEQ ID NO.: 35).

Following amplification of the triple helix-forming target sequence (by the methods described in the above examples), a triplex probe which includes a sequence complementary to the duplex PCR product would be used to detect the presence of the organism. For example, an HIV-1 triplex probe may include the sequence complementary to

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nucleotides 404-419 of HIV-1 (i.e., tccccctttctttttt-NH₂; SEQ ID NO.: 36).

Other Embodiments

Other embodiments are within the following claims. For example, any target sequence containing a triple helix-forming sequence (either naturally or as a result of some amplification process) may be detected by these methods. The target sequence may be DNA or RNA. Detection may be accomplished by any reporter group, for example, any detectable fluorescent, enzymatic, radioactive, chemiluminescent, or bioluminescent reporter group. Alternatively, aggregation-based, mass-based, interference-based or absorption-based reporter groups may be used to capture and/or detect triple helical product. Capture and/or detection of these reporter groups is well known to those skilled in the art

The methods of the invention may be carried out using assay formats other than a microtiter well or an FCA plate. For example, mass-based surface acoustic wave or surface transverse wave techniques, fluorescence analysis using fiber optic wave guides, evanescent zone analysis, or any other format which allows detection of the triple helical product may be used. Moreover, the process may be automated, for example, by using an PCFIA plate format.

As described above, detection of target sequences is accomplished using triple helix probes that permit capture and detection. Capture and detection may be accomplished by separate probe molecules, each hybridizing at a different site (i.e., a capture probe and a detection probe), in which case the "third strand" of the invention includes two separate molecules. Alternatively, both these functions

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may be achieved with a single triplex probe. In one particular example, a single probe is bound at one end to a Latex microparticle (for capture) and at the other end to sulforhodamine (for detection). In addition, we have shown that triple helix formation increases the ultraviolet fluorescence of ethidium bromide intercalated into the duplex target sequence. When triple helical formation is assayed as an increase in ethidium bromide fluorescence, there is no need to covalently bind a reporter molecule to a triplex probe.

Triple helix formation may also be measured by a competitive binding assay. In one example, product duplexes (generated as described above) containing a triple helix-forming region are hybridized to a third strand triplex probe to which is covalently bound a detectable label, e.g., sulforhodamine. To the hybridization mixture is then added a single-stranded or double-stranded "hook" sequence to which is covalently bound a solid support, e.g., a Latex microparticle. The hook sequence is complementary to the labelled third strand triplex probe. Triplex probe sequences which bind to the solid support-bound hook sequence (i.e., those third strands which are in excess following hybridization with product duplexes) are isolated (e.g., on an FCA plate following vacuum filtration or by centrifugation) and assayed (e.g., by measuring sulforhodamine fluorescence). The presence and/or concentration of product duplexes is inversely proportional to the quantity of label detected. Such capture of excess triplex probe by hook sequence is facilitated by triple helical, e.g., Hoogsteen, interactions in the case of the double-stranded hook and double helical, e.g., Watson-Crick, interactions in the case of the single-stranded hook sequences.

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The methods of the invention allow detection of any target sequence (i.e., from any organism) whose sequence, or a portion thereof, has been determined. For example, pathogenic organisms, including, but not limited to, retroviruses (e.g., human immunodeficiency virus-1, feline leukemia virus, feline immunodeficiency virus), hepatitis virus, bovine viral diarrhea virus, and papilloma viruses (e.g., human papilloma viruses, type 16 and type 18) may be detected by designing appropriate triplex primers and probes (based on published sequences) and assaying for such sequences as described above.

Fig. 8 is a list of genes which include putative endogenous triple helix-forming sequences. The genes were obtained from the GENBANK database based upon their possession of a perfect 25 base pair homopolymer region. The first column lists the GENBANK Accession Code, and the second column lists the full name of the gene and the organism from which it derives. Using this approach, other homopolymer-containing genes may be identified; accessing the full sequence of such genes would allow the exact homopolymer sequences and their positions within the genes to be determined. Fig. 8 demonstrates that homopolymer tracts, i.e., endogenous triple helix-forming sequences, are resident in the genome of a variety of organisms and provide candidate sequences for detection by the methods of this invention.

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COMPUTER SUBMISSION OF DNA AND AMINO ACID SEQUENCES

(1) GENERAL INFORMATION:

(i) APPLICANT: Vary, Calvin

(ii) TITLE OF INVENTION: NUCLEIC ACID SEQUENCE
DETECTION BY TRIPLE HELIX
FORMATION

(iii) NUMBER OF SEQUENCES: 50

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Fish & Richardson
(B) STREET: One Financial Center
(C) CITY: Boston
(D) STATE: Massachusetts
(E) COUNTRY: U.S.A.
(F) ZIP CODE: 02111-2658

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: 3.5" Diskette, 1.44 Mb storage
(B) COMPUTER: IBM PS/2 Model 50Z or 55SX
(C) OPERATING SYSTEM: IBM P.C. DOS (Version 3.30)
(D) SOFTWARE: WordPerfect (Version 5.0)

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: N/A
(B) FILING DATE: N/A
(C) CLASSIFICATION: N/A

(vii) PRIOR APPLICATION DATA:

SUBSTITUTE SHEET

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Prior applications total,
including application
described below: None

(A) APPLICATION NUMBER: N/A
(B) FILING DATE: N/A

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Freeman, John W.
(B) REGISTRATION NUMBER: 29,066
(C) REFERENCE/DOCKET #: 00088-037001

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (617) 542-5070
(B) TELEFAX: (617) 542-8906
(C) TELEX: 200154

TOTAL NUMBER OF SEQUENCES TO BE LISTED: 50

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 1

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 1

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 1

GGAAGGAAAG AAGGAG

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(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 2

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 2

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 2

CCTTCCTTTC TTCCTC

16

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 3

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 3

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 3

CCTTCCTTTC TTCCTC

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(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 4

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 39
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 4

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 4

CCCCCTTTTTC TCTCCTTTCT CCGACAATGA CGGTACGG 39

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 5

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 38
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 5

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 5

CCTCTCTTCT CTCTTCTCTC GGACAATGAC GGTTACGG 38

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 6

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 41
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 6

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 6

CCCCTTTTTC TCTCCTTTCT CCAAGGCGAT CAGCAACGCG G 41

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 7

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 19
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 7

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 7

CGGACAATGA CGGTTACGG 19

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 8

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single

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(D) TOPOLOGY: linear

(E) NAME: SEQ ID NO.: 8

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 8

CAAGGCGATC AGCAACGCGG

20

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 9

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(E) NAME: SEQ ID NO.: 9

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 9

CTCTTTCCTC TCTTTTCCC C

21

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 10

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(E) NAME: SEQ ID NO.: 10

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(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 10

CTCTCTTCTC TCTTCTCTCC

20

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 11

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 11

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 11

GCCCCGAACG CCGATACC

18

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 12

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 12

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 12

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CCCAGGATGA CGCCGAAT

60

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 13

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 39
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 13

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 13

CCCCTTTTTC TCTCCTTTCT CGCCGCTAAC GCCCAACAC 39

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 14

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 38
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 14

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 14

CCTCTCTTCT CTCTTCTCT CGCTCCTCG ATCATCGC 38

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(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 15

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 37
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 15

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 15

CGAATCAAAT CAAACTAACC CCTTTTCTC TCCTTTC 37

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 16

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 18
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 16

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 16

TCTAACTCTG TCATCATC 18

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 17

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 21
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 17

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 17

TTCTTCTTCT TCTTTCTTCT T 21

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 18

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 18

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 18

TAGGAAAGGC ACCCCCACAT TGG 60

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 19

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21
(B) TYPE: nucleic acid

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(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 19

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 19

CCCCTAAGAT CTCCTCCATG G

21

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 41
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 20

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 20

CCCCTTTTTC TCTCCTTCT CATTAGGACT TCCGCATCCG G

41

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 21

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 41
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 21

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(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 21

CCTCTCTTCT CTCTTCTCTC GCAAGTGTAC TCTCGATATG G 41

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 22

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 22

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 22

GCACTTTAAA TTTTCCCAT T AGTCC 25

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 23

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 23

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 23

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CCTGCGGGAT GTGGTATTCC

20

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 24

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 40
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 24

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 24

CCCCTTTTTC TCTCCTTTCT CAAGCCAGGA ATGGATGGCC 40

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 25

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 39
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 25

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 25

CCTCTCTTCT CTCTTCTCTC CAGAAGTCTT GAGTTCTCC 39

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(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 26

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 26

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 26

AGGGGGAAAG AAAAAA 16

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 27

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 27

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 27

AAATGGAAAA GGAAGGGAAA A 21

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 28

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 15
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 28

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 28

AAGGGAAAAA GGAAA

15

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 29

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 33
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 29

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 29

AAAAGGGAGA CCCCAGAGGA AAGGGAAGAA AGA

33

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 30

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 24
(B) TYPE: nucleic acid

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(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 30

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 30

AGAGGAGGAG GATGAAATAG ATGG

24

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 31

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 11
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 31

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 31

GGGGAAGAGG G

11

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 32

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 32

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(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 32

AAAAATAGAT GAAGGGGGAG A

21

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 33

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 14
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 33

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 33

GAGGAAGAGG AAGA

14

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 34

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 34

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 34

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TCCCCTTCTT CTTCTGCCGT TCC

23

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 35

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 35

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 35

CTCCCTCCTT TCCTC

15

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 36

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 36

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 36

TCCCCCTTTC TTTTTT

16

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(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 37**(i) SEQUENCE CHARACTERISTICS:**

(A) LENGTH: 16
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 37

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 37

TCCCCCTTTC TTTTTT

16

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 38**(i) SEQUENCE CHARACTERISTICS:**

(A) LENGTH: 21
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ. ID NO.: 38

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 38

TTTACCTTTT CCTTCCCTTT T

21

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 39**SUBSTITUTE SHEET**

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(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 39

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 39

TTCCCTTTT CTTT

15

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 40

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 33
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 40

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 40

TTTCCCTCT GGGTCTCCT TTCCCTTCTT TCT

33

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 41

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21

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(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 41

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 41

TTCTTCTTCT TCTTTCTTCT T

21

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 42

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 37
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 42

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 42

TTTTCTTTT CTGTTTCTTC TTCTTTCTTT CTTCTTT

37

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 43

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 11
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(E) NAME: SEQ ID NO.: 43

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 43

CCCCTTCTCC C

11

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 44

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 24
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 44

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 44

TCTCCTCCTC CTACTTTATC TACC

24

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 45

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 45

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 45

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TTTTTATCTA CTTCCCCCTC T

21

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 46

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 14
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 46

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 46

CTCCTTCTCC TTCT

14

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 47

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 47

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 47

AGGGGAAGAA GAAGACGGCA AGG

23

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(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 48**(i) SEQUENCE CHARACTERISTICS:**

(A) LENGTH: 15
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 48

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 48

GAGGGAGGAA AGGAG

15

(2) INFORMATION FOR SEQUENCE IDENTIFICATION NUMBER: 49**(i) SEQUENCE CHARACTERISTICS:**

(A) LENGTH: 37
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(E) NAME: SEQ ID NO.: 49

(ii) SEQUENCE DESCRIPTION FOR SEQUENCE ID NUMBER: 49

AAAAGGAAAA GACAAAGAAG AAGAAAGAAA GAAGAAA

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Claims

1 1. A method for detecting a nucleic acid, said
2 method comprising the steps of:
3 amplifying said nucleic acid in vitro using cycles
4 of denaturation and amplification to yield product duplexes,
5 and
6 detecting one of said product duplexes by
7 hybridizing a third strand of nucleic acid to said product
8 duplexes without denaturation.

1 2. The method of claim 1, wherein said amplifying
2 is accomplished by polymerase chain reaction.

1 3. The method of claim 2, wherein said polymerase
2 chain reaction is accomplished using at least two PCR
3 primers, one of said PCR primers comprising a sequence
4 hybridizable with said nucleic acid and one strand of a
5 triple helix-forming sequence, the other primer being
6 hybridizable to the complement of said nucleic acid.

1 4. The method of claim 1, wherein said amplifying
2 is accomplished by ligase chain reaction.

1 5. The method of claim 4, wherein said ligase chain
2 reaction is accomplished using two or more pairs of LCR
3 hybridization probes, wherein one said probe of each pair
4 comprises a sequence hybridizable to said nucleic acid and
5 the second said probe of each pair comprises both a sequence
6 complementary to an adjacent segment of said nucleic acid
7 and a sequence comprising one strand of a triple helix-
8 forming sequence.

1 6. A method for detecting the presence of a
2 pathogen in a biological sample, said pathogen comprising an
3 endogenous triple helix-forming nucleic acid sequence, said
4 method comprising hybridizing a third strand of nucleic acid
5 to a duplex comprising said endogenous triple helix-forming
6 sequence without denaturation of said duplex.

1 7. The method of claim 1 or 6, wherein said third
2 strand of nucleic acid comprises a polypyrimidine sequence
3 of at least 15 nucleotides.

1 8. The method of claim 7, wherein said third strand
2 of nucleic acid further comprises one or more purine
3 residues, wherein each said purine residue is flanked by 9
4 or more pyrimidine residues.

1 9. The method of claim 1 or 6, wherein said third
2 strand of nucleic acid comprises a polypurine sequence of at
3 least 15 nucleotides.

1 10. The method of claim 9, wherein said third
2 strand of nucleic acid further comprises one or more
3 pyrimidine residues, wherein each said pyrimidine residue is
4 flanked by 9 or more purine residues.

1 11. The method of claim 7 or 9, wherein said
2 polypyrimidine or polypurine sequence comprises one or more
3 modified nucleotides.

4 12. The method of claim 1 or 6, wherein said third
5 strand of nucleic acid is covalently attached to a solid
6 support.

1 13. The method of claim 12, wherein said solid
2 support is a microparticle.

1 14. The method of claim 1 or 6, wherein said
2 nucleic acid is detected using an FCA format.

1 15. The method of claim 1 or 6, wherein said
2 nucleic acid is detected using a PCFIA format.

1 16. The method of claim 1 or 6, wherein said
2 nucleic acid is detected using a microtiter well format.

1 17. The method of claim 1 or 6, wherein said third
2 strand of nucleic acid is covalently attached to a reporter
3 group.

1 18. The method of claim 17, wherein said reporter
2 group is sulforhodamine.

1 19. The method of claim 17, wherein said reporter
2 group is alkaline phosphatase.

1 20. The method of claim 1 or 6, wherein said
2 nucleic acid is detected by enhanced ethidium bromide
3 fluorescence following contact with said third strand.

1 21. The method of claim 1 or 6, further comprising
2 isolating said product prior to detecting said product.

1 22. The method of claim 1 or 6, further comprising
2 detecting a second nucleic acid.

1 23. The method of claim 22, wherein said detection
2 of said second nucleic acid duplex, different from said
3 product duplexes, by a method comprising hybridizing said
4 second duplex a third strand of nucleic acid.

1 24. The method of claim 1, wherein said nucleic
2 acid is derived from M. paratuberculosis.

1 25. The method of claim 1 or 6, wherein said
2 nucleic acid is derived from a retrovirus.

1 26. The method of claim 25, wherein said retrovirus
2 is a human immunodeficiency virus.

1 27. The method of claim 26, wherein said third
2 strand comprises at least 15 consecutive nucleotides of the
3 sequence: tccccctttctttttt (SEQ ID NO.: 37).

1 28. The method of claim 26, wherein said third
2 strand comprises at least 15 consecutive nucleotides of the
3 sequence: ttaccttttccttcctttt (SEQ ID NO.: 38).

1 29. The method of claim 25, wherein said retrovirus
2 is a feline leukemia virus.

1 30. The method of claim 29, wherein said third
2 strand comprises the sequence: ttccctttttccttt (SEQ ID NO.:
3 39).

1 31. The method of claim 29, wherein said third
2 strand comprises at least 15 consecutive nucleotides of the
3 sequence: ttttcctctggggtctcctttcccttctttct (SEQ ID NO.:
4 40).

1 32. The method of claim 25, wherein said retrovirus
2 is a feline immunodeficiency virus.

1 33. The method of claim 32, wherein said third
2 strand comprises at least 15 consecutive nucleotides of the
3 sequence: ttcttcttcttctttcttctt (SEQ ID NO.: 41).

1 34. The method of claim 32, wherein said third
2 strand comprises at least 15 consecutive nucleotides of the
3 sequence: ttttccttttctgtttcttcttctttctttcttctt (SEQ ID NO.:
4 42).

1 35. The method of claim 1 or 6, wherein said
2 nucleic acid is derived from a human papilloma virus.

1 36. The method of claim 35, wherein said human
2 papilloma virus is HP-16.

1 37. The method of claim 36, wherein said third
2 strand comprises the sequence: ccccttctccc (SEQ ID NO.:43).

1 38. The method of claim 36, wherein said third
2 strand comprises at least 15 consecutive nucleotides of the
3 sequence: tctcctcctcctactttatctacc (SEQ ID NO.: 44).

1 39. The method of claim 35, wherein said human
2 papilloma virus is HP-18.

1 40. The method of claim 39, wherein said third
2 strand comprises at least 15 consecutive nucleotides of the
3 sequence: tttttatctacttccccctct (SEQ ID NO.: 45).

1 41. The method of claim 39, wherein said third
2 strand comprises the sequence: ctccttctccttct (SEQ ID NO.:
3 46).

1 42. The method of claim 25, wherein said retrovirus
2 is a caprine arthritis encephalitis virus.

1 43. The method of claim 1 or 6, wherein said
2 nucleic acid is derived from a hepatitis B virus.

1 44. The method of claim 43, wherein said third
2 strand comprises at least 15 consecutive nucleotides of the
3 sequence: aggggaagaagaagacggcaagg (SEQ ID NO.: 47).

1 45. The method of claim 43, wherein said third
2 strand comprises the sequence: gagggaggaaaggag (SEQ ID NO.:
3 48).

1 46. A purified single-stranded nucleic acid probe
2 comprising at least 15 consecutive nucleotides of the triple

3 helix-forming sequence: ctctttcctctcttttttcccc (SEQ ID NO.:
4 9).

1 47. A purified single-stranded nucleic acid probe
2 comprising at least 15 consecutive nucleotides of the triple
3 helix-forming sequence: ctctcttctctcttctctcc (SEQ ID NO.:
4 10).

1 48. A PCR primer comprising a sequence hybridizable
2 with a target sequence nucleic acid and one strand of a
3 triple helix-forming sequence.

1 49. An LCR hybridization probes, wherein said probe
2 comprises a sequence hybridizable to a target sequence
3 nucleic acid and a sequence comprising one strand of a
4 triple helix-forming sequence.

1 50. A kit for detecting a nucleic acid, said kit
2 comprising a single-stranded nucleic acid probe capable of
3 specifically hybridizing to a triple helix-forming sequence
4 of said nucleic acid, said probe being covalently attached
5 to a reporter group, and
6 a reagent for detecting said reporter group bound to
7 said nucleic acid.

1 51. The kit of claim 50, further comprising a
2 second single-stranded nucleic acid capable of specifically
3 hybridizing to a triple helix-forming sequence of said
4 nucleic acid, wherein said second single-stranded nucleic
5 acid is covalently attached to a solid support.

1 52. The kit of claim 50, wherein said single-
2 stranded nucleic acid is covalently attached to a solid
3 support.

1 53. The kit of claim 51 or 52, wherein said solid
2 support is a microparticle.

1 54. The kit of claim 50, wherein said nucleic acid
2 is detected using a microtiter well format.

1 55. The kit of claim 50, wherein said nucleic acid
2 is detected using an FCA format.

1 56. The kit of claim 50, wherein said nucleic acid
2 is detected using a PCFIA format.

1 57. The kit of claim 50, wherein said reporter group
2 is alkaline phosphatase.

1 58. The kit of claim 50, wherein said reporter
2 group is sulforhodamine.

1 59. The kit of claim 50, further comprising at
2 least two PCR primers, one of said PCR primers comprising a
3 sequence hybridizable with said nucleic acid and one strand
4 of a triple helix-forming sequence, the other primer being
5 hybridizable to the complement of said nucleic acid.

1 60. The kit of claim 50, further comprising two or
2 more pairs of LCR hybridization probes, wherein one said
3 probe of each pair comprises a sequence hybridizable to a

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4 target sequence nucleic acid and the second said probe of
5 each pair comprises both a sequence complementary to an
6 adjacent segment of said target sequence nucleic acid and a
7 sequence comprising one strand of a triple helix-forming
8 sequence.

1 61. A method for detecting a nucleic acid, said
2 method comprising the steps of:
3 amplifying said nucleic acid in vitro using cycles
4 of denaturation and amplification to yield product duplexes,
5 hybridizing a third strand of nucleic acid to said
6 product duplexes without denaturation,
7 hybridizing any free said third strand with a
8 nucleic acid hook sequence covalently bound to a solid
9 support, and
10 measuring the amount of said product duplex as the
11 inverse measure of the quantity of said third strand bound
12 to said hook sequence.

1 62. The method of claim 61, wherein said nucleic
2 acid hook sequence is double-stranded.

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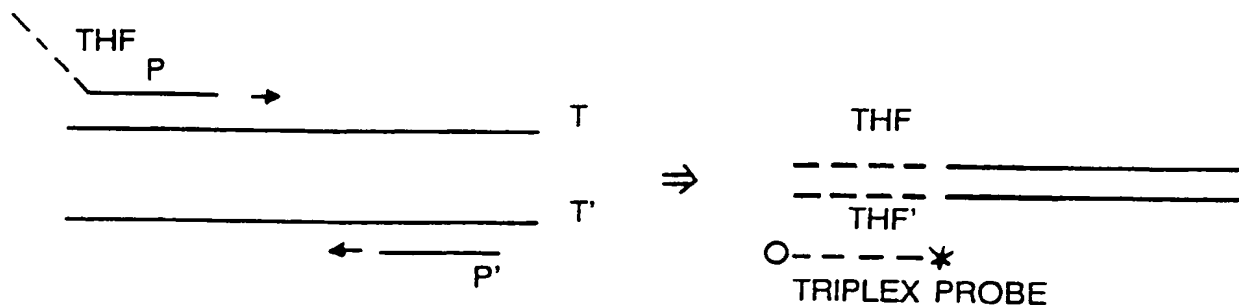


FIG. 1

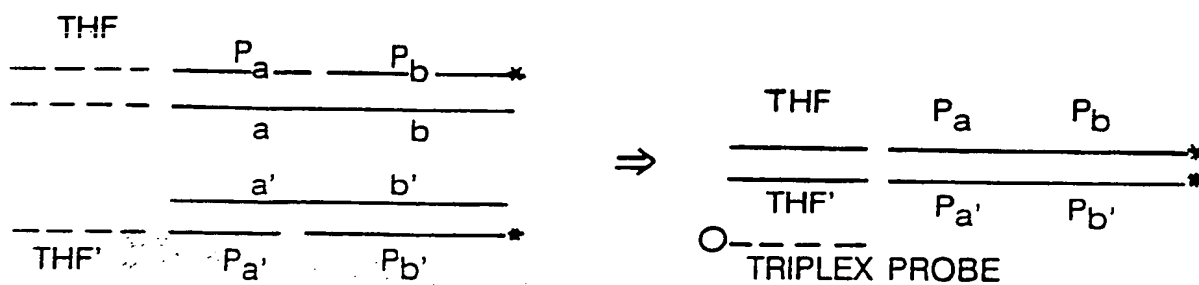


FIG. 2

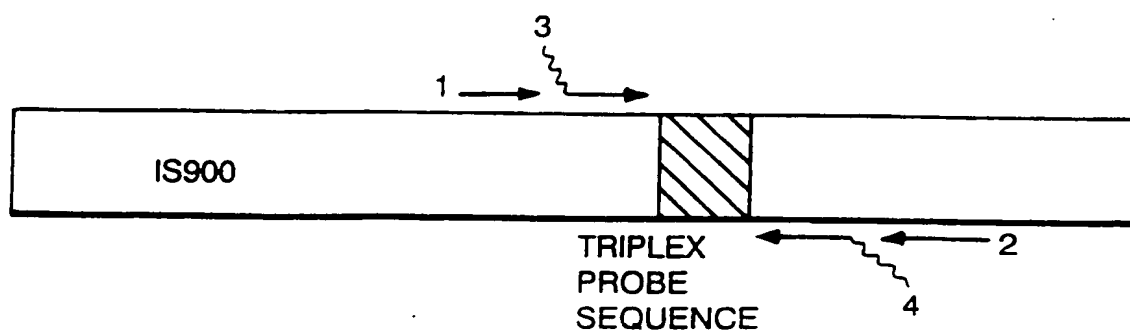


FIG. 3

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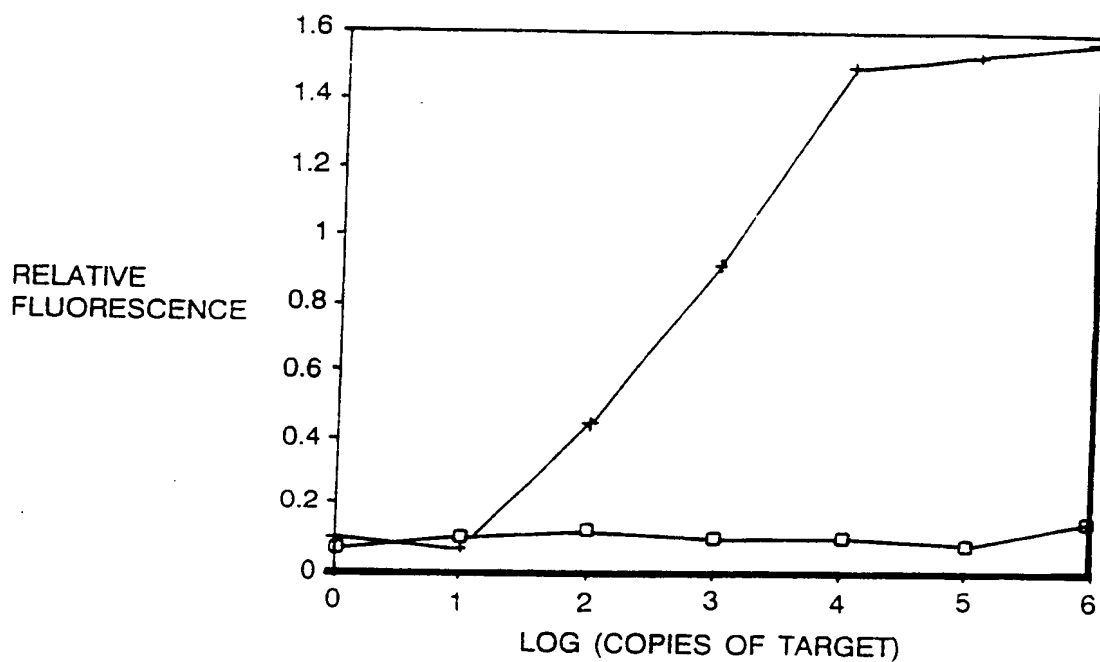


FIG. 4

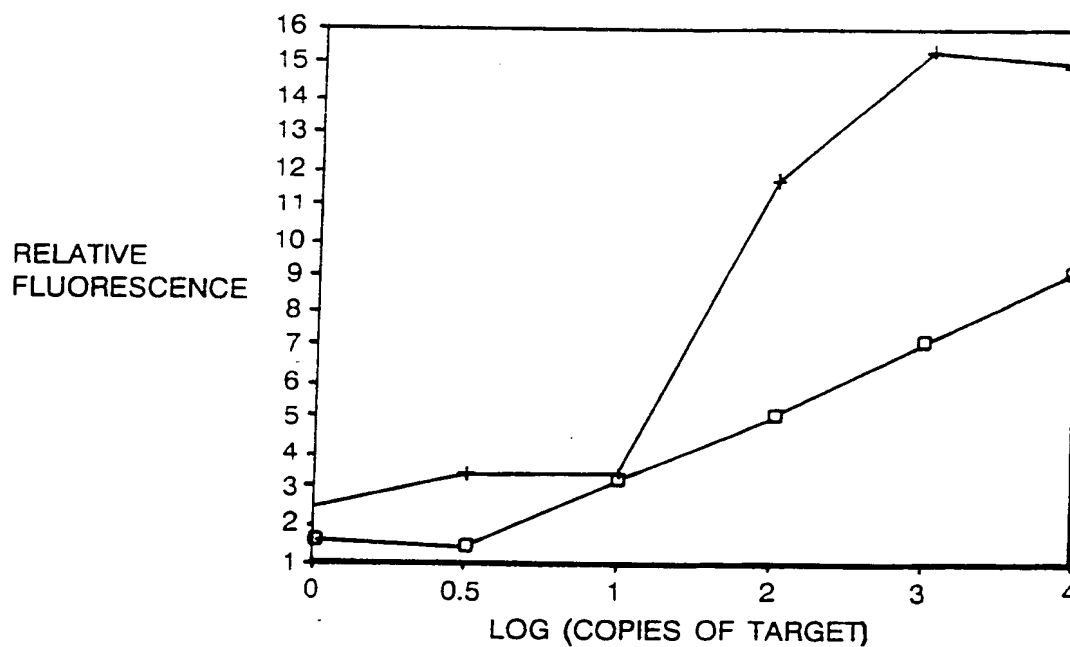


FIG. 5

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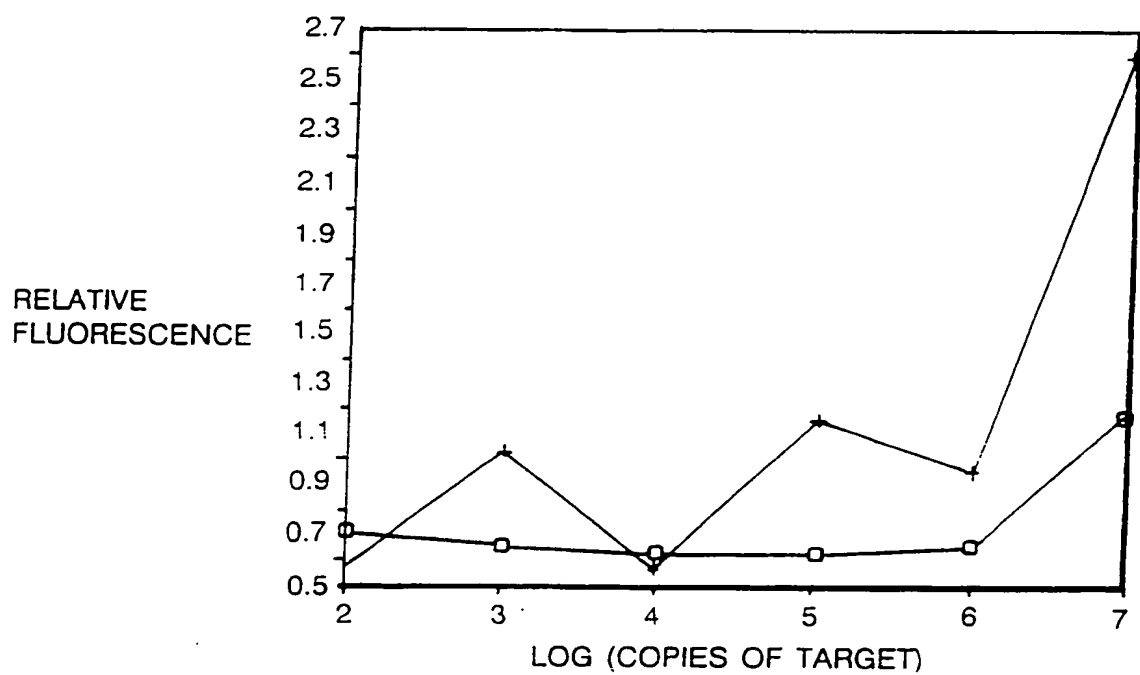


FIG. 6

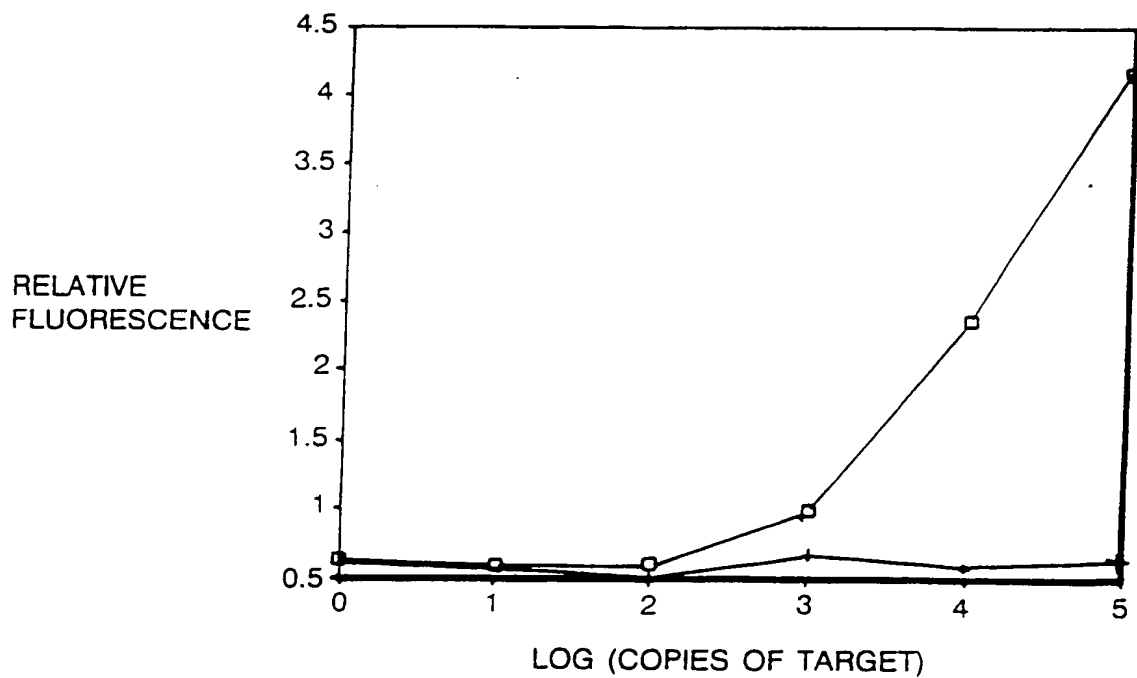


FIG. 7

<u>GENBANK</u> <u>CODE</u>	<u>GENE</u>
HUMGCB1	EHuman glucocerebrosidase gene, complete cds.
HUMGASTA	EHuman gastrin gene, complete cds.
HUMGHCSA	EHuman growth hormone (GH-1 and GH-2) and chorionic
HUMFOLLI1	EHuman follistatin gene, exons 1-5.
HUMRBA	EHuman retinoblastoma (pp110RB) gene, 5' flank.
HUMRASR2	EHuman R-ras gene, exons 2 through 6.
HUMFOLLI2	EHuman follistatin gene, exon 6.
HUMPKCB2A	EHuman mRNA for protein kinase C (PKC) type beta II
HUMGP5MOS	EHuman gene fragment related to oncogene c-mos with
HUMGSHPXG	EHuman glutathione peroxidase gene, complete cds.
HUMARASR1	EHuman R-ras gene, exon 1.
HUMPP15	EHuman gene for PP15 (placental protein 15).
HUMPRCM	EHuman protein C mRNA, complete cds.
HUMCYPIIE	EHuman cytochrome P450IIE1 (ethanol-inducible) gene
HUMERP	EHuman erythropoietin gene, complete cds.
HUMCYCPSJ	EHuman somatic cytochrome c (HS11) processed pseudo
HUMFCERI	EHuman mRNA for high affinity IgE receptor alpha-su
HUMPLAP1A	EHuman placental alkaline phosphatase (PLAP-1) gene
HUMERPA	EHuman erythropoietin gene, complete cds.
HUMIFNIN3	EHuman interferon-inducible mRNA fragment (cDNA 6-1
HUMIFNB3	EHuman interferon-beta-3 gene.
HUMMHDDR3	EHuman MHC class II lymphocyte antigen DPw4-beta-2
HUMIGHAD	EHuman Ig rearranged H-chain epsilon-3 pseudogene,
HUMMHDC3B	EHuman MHC class II HLA-DC-3beta gene (DR3,3).
HUMMRP8A	EHuman migration inhibitory factor-related protein
HUMHSC70	EHuman hsc70 gene for 71 kd heat shock cognate prot
HUMMLVI2	EHuman genomic Mlvi-2 locus with Alu insert.
HUMMPOA	EHuman myeloperoxidase gene, exons 1-4.
HUMKAL1	EHuman glandular kallikrein gene, region 5' to the
HUMINSCR	EHuman insulin receptor (allele 1) gene, exons 14,
HUMKEREP	EHuman 50 kd type I epidermal keratin gene, complet
HUMKER18	EHuman keratin 18 (K18) gene, complete cds.
HUMKERP2	EHuman keratin pseudogene, exons 2-8.
HUMINCPPS	EHuman cysteine-pseudogene, inhibitor pseudogene (CS
HUMHSHXH	EHuman MHC class II HLA-SX-alpha gene.
HUMINSR	EHuman insulin receptor mRNA, complete cds.
HUMMAC1A	EHuman Mac-1 gene encoding complement receptor type
HUMP5311	EHuman cellular phosphoprotein p53 gene, exon 11.
HUMHBBBP1	EHuman beta-globin gene cluster, region homologous
HUMP13093	EHuman mRNA for leukocyte adhesion glycoprotein p15
HUMHBB	EHuman beta globin region on chromosome 11.
HUMHBB	EHuman beta globin region on chromosome 11.
HUMHBB51	EHuman beta globin gene cluster extreme 5' flank: d
HUMHBB	EHuman beta globin region on chromosome 11.
HUMHOXB	EHuman homeo box c8 protein, mRNA, complete cds.
HUMNGLB	EHuman beta-nerve growth factor (beta-NGF) gene.
HUMMYCC	EHuman (Lawn) c-myr proto-oncogene, complete coding

FIG. 8a

<u>GENBANK CODE</u>	<u>GENE</u>
HUMMYCRT	EHuman (Raji) translocated t(8;14) c-myc uncogene,
HUMINSTR	EHuman insulin receptor mRNA, complete cds.
HUMNMYCA	EHuman germ line n-myc gene.
HUMNMYC	EHuman N-myc gene. exons 8 and 3.
PIGPKCAMP	EPig CAMP-dependent protein kinase catalytic alpha
PIGMHDRG	ESwine MHC class I PDE-glycoprotein mRNA, complete
RABBGLOB	ERabbit beta-like globin gene cluster encoding the
PIGFSA	EPig follistatin (FS) gene, complete cds.
RABBGLOB	ERabbit beta-like globin gene cluster encoding the
GOTHBBPS1	EGoat beta-x-globin pseudogene with 3' flank.
BOVCOXPS	EBovine cytochrome c oxidase subunit IV processed p
DOGGPCR8	EC.familiaris mRNA for G protein-coupled receptor,
RABMHDP	ERabbit MHC class II DP alpha-1 gene (RLA K10 hapl
RABPHXC	ERabbit phosphatase X catalytic subunit mRNA, 3' en
PIGRELXA	EPorcine relaxin gene, complete cds.
BOVHMG1	EBovine mRNA for high mobility group 1 (HMG1) prote
GOTHBBZPS	EGoat beta-z-globin pseudogene.
DOGGPCR1	EC.familiaris mRNA for G protein-coupled receptor,
BOVTDTR	EBovine mRNA for terminal deoxynucleotidyltransfera
RABBGLOB	ERabbit beta-like globin gene cluster encoding the
RABBGLOB	ERabbit beta-like globin gene cluster encoding the
SHPCRFA	ESheep corticotropin releasing factor gene, complet
RABUTGLOS	ERabbit uteroglobin gene 5'-flank EcoRI2-EcoRI3 fra
PIGAPOB2	EPig apolipoprotein B gene (Lpb), exons 26 through
RABHBB1B1	ERabbit beta1-globin gene (allele 2), complete cds
BOVPRCI3	EBovine placental prolactin-related protein (bPRC-I
BOVTMD	EBovine thrombomodulin mRNA, 3' end.
PIGTFR	EPorcine transferrin mRNA, 3' end.
RABRSCA	ERabbit short interspersed C repeat (SINE), about 1
PININHBAR	EPorcine mRNA for inhibin beta (a)-subunit.
PIG2APHA	EPorcine protein phosphatase 2A alpha subunit mRNA,
BOVPGII	EBovine mRNA for bone proteoglycan II.
BOVATPS	EBovine mitochondrial ATP synthase gamma subunit ge
BOVMARCKS	EBovine 80-87 kd myristoylated alanine-rich C kinas
DOGATPBR	EDog kidney mRNA for (Na ⁺ /K ⁺)-ATPase beta-subunit.
RABSTROMR	ERabbit stromelysin gene, 5' flank.
GOTHBBEII	EGoat embryonic beta-globin epsilon-II complete gen
BOVPTHG	EBovine parathyroid hormone gene, complete coding r
BOVTHYR5	EBovine thyroglobulin gene exon 18.
BOVGLYAA3	EBovine pituitary glycoprotein hormone alpha-subuni
BOVKERVIC	EBovine epidermal cytokeratin VIIb gene, complete cd
LEEBGLOB	ELepus europaeus adult beta-globin gene.
BOVPKC	EBovine beta type protein kinase C mRNA, complete c
BOVIRBP	EBovine interphotoreceptor retinoid-binding protein
RABDPG	ERabbit reticulocyte 2,3-bisphosphoglycerate (DPG)
BOVHBBE4	EBovine epsilon-4 beta-globin gene, complete cds.
BOVHBP1	EBovine beta-globin psi-1 pseudogene, complete cds.
BOVHBP2	EBovine beta-globin psi-2 pseudogene, complete cds.

FIG. 8b

<u>GENBAND</u> <u>CODE</u>	<u>GENE</u>
CHKCPG	EChicken proteoglycan core protein n= gene lase 5 exo
CHKPPGA	EChicken processed pseudogene, complete cds.
CHKCPS1	EChicken processed pseudogene CPS1 related to the r
CHKOVAL	Echicken ovalbumin gene including flanking sequence
CHKCRYAA	EChicken alpha-A-crystallin gene, complete cds and
CHKMYLCC	EChicken myosin alkali light chain (MLC1-f/MLC3-f)
CHKCG1A1	EChicken pro-alpha-1 collagen(I) mRNA, 3'end.
CHKCERBA1	EChicken c-erb A gene exon 1 and flanks.
CHKCG1A2	EChicken type-1 collagen pro-alpha-2, exon 2 & 3.
CHKCMYCA	EChicken tumor 10 c-myc DNA, exons 2 ad 3.
CHKTGFB4	EChicken mRNA for transforming growth factor-beta 4
CHKMHC3	EChicken embryonic myosin heavy chain (MHC) gene 3'
CHKMLC131	EChicken fast myosin alkali light chain, exon 1 spe
CHKC1G	EChicken histone H1 gene, clone lambda-CHQ1.
CHKH2A	EChicken histone H2A gene.
CHKH234G	EChicken histone LH4, RH4, H3, LH2A and RH2A genes.
CHKH2A2B	EChicken histone H2A/H2B gene pair and flanks.
CHKMYHE	EChicken embryonic myosin heavy chain gene, complet
CHKCOACA	EChicken acetyl-CoA carboxylase mRNA, complete cds.
CHKCYP450	EChicken cytochrome P-450 (phenobarbital-inducible)
CHKDYS	EChicken mRNA for dystrophin (Suchenne muscular dys
CHKMYHB	EChicken fast-white myosin heavy chain (adult isofo
XELH1PS1	Exenopus laevis h1 histone pseudogene.
XELHI53LA	Exenopus laevis h3 histone mrna.
XEBACTA3	EX.borealis cytoskeletal actin type 1 gene, exon 4.
XELCONNEX	EX.laevis connexin 38 mRNA, complete cds.
CHKC1PA1M	EChicken type-1 collagen pro-alpha-1 chain mRNA.
APTRDNA	EAscapus truei ribosomal DNA intergenic spacer.
XELHX1H3	EX.laevis histone H1B, H2A, H2B, and H4 genes, comp
XELSEKIIB	EX.laevis ribosomal protein S6 kinase II bata (SEKI
XELHX1H1	EX.laevis histone H1B, H2A, H2B, and H4 genes, comp
DUKHGAP	EDuck embryonic alpha-globin pi' gene, complete cds.
DUKCRYD3A	EDuck delta-crystallin gene exon 3 fragment.
CHKTNC	EChicken troponin C (TNC) mRNA, complete cds.
CHKVITUP	EChicken vitellogenin gene upstream region (2kb).
CHKY	Echicken y gene, including flanking wequences.
DUKHBB	EDuck beta-globin mrna.
CHKMYC	EChicken cellular myc proto-oncogene, complete cds.
DUKH5	EDuck (Cairina moschata) H5 histone gene, complete
CHKTHD	EChicken thioredoxin protein, complete cds.
CHKLNKPG	EChicken cartilage link protein mRNA, complete cds.
CHKFASA	EChicken fatty acid synthase gene, 3' and.
RANFERPG	EFrog (Rans catesbelana) apoferritin pseudogene.
CHKFERH	EChicken ferritin H-subunit gene, complete cds.
DUKFASA	EDuck (A.platyrnynchos) S-acyl fatty acid synthase
CHKMLC138	EChicken fast myosin alkali light chains, exons 1 a
ONGRAL1A	EOncnocer volvunus RAL-1 mRNA, complete cds.
OCTHEM	EO.dorlein memocyanin mRNA, 3' end.
TRBRGAB	ET.brucei trucei 5S ribosomal RNA gene, clone p5S-2

FIG. 8c

<u>GENBANK</u> <u>CODE</u>	<u>GENE</u>
TRBRGAC	ET.brucei rhodesiense 5S ribosomal RNA gene, clone
DDIMYHC	ED.discoideum myosin heavy chain gene, complete cds
TRBRGAA	ET.brucei brucei 5S ribosomal RNA gene, clone p5S-1
NEMRDNA2	EAscaris bumbricoides rDNA with 18S rRNA 5' end.
DDIRAS	ESlime mold (D.discoideum) ras-homologous gene, com
DDIPSTCAT	EDictyostelium discoideum pst-cath gene encoding ps
DDIPYR56G	EDictyostelium discoideum DdPYR5-6 gene for UMP syn
DDIHPERB	ED.discoideum protein 23Cgene, 5' end.
DDIDIFIND	ED.discoideum DIF-inducible mRNA.
PFASA7	EPlasmodium falciparum (isolate NF7) S antigen gen
PFASA27	EPlasmodium falciparum (isolate FC27) S-antigen gen
DDICYSPRO	ESlime mold (D.discoideum) cysteine proteinase 1 mR
DDIDG17A	EDictyostelium discoideum DG17 gene, complete cds.
DDIDRE1AC	ED.discoideum insertion elements DRE1a and DRE2-III
TRBRGAD	ET.brucei 5S ribosomal gene, complete cds.
DDIDOD	ESlime mold (D.discoideum) dihydroorotate dehydroge
DDIDISCIG	ESlime mold (D.discoideum) discoidin I-gamma gene,
DDIDISIA	ESlime mold (D.discoideum) discoidin-ia gene.
PBSCSP	EP.brasilianum circumsporozoite protein gene, 3'en
PFAP41R	EP.falciparum aldolase (P41) gene, complete cds.
SCMHGPRT	ES.mansoni hypoxanthine-guanine phosphoribosyltrans
DROTNF192	ED.melanogaster transposable element F19, 3' juncti
PLMCSP	EP.malariae circumsporozoite protein gene, complete
PFARESAG1	EP.falciparum FC27 RESA gene for ring-infected eryt
DROLAMB1	ED.melanogaster laminin B1 subunit mRNA, complete c
DROORFI	EDrosophila DNA for hybridizing with human preproin
SUSHISPS3	ESea urchin (S.purpuratus) early histone H2Bpseudo
DROGBR	ED.melanogaster glue gene cluster 68C boundary regi
DROH2AVD	ED. melanogaster H2AvD mRNA for histone 2A variant.
PFARESAR1	EP.falciparum FC27 Ag46 RESA mRNA for ring-infected
DDITGVM	ED.discoideum Val-tRNA gene, clone lambda-ValGUU13.
DDITND312	ESlime mold (D.discoldeum strain Ax-3L) transposons
NEM18SRN5	EAscaris lumbricoides 12S ribosomal DNA 5" region.
DDITGVF	ED.discoideum Val-tRNA gene, clone lambda-ValGUUB.
DDITGVH	ED.discoldeum val-tRNA gene, clone lambda-ValGUUB.
DROYPI2	ED.meianogaster ypl and yp2 genes, encoding yolk pr
DROTPOD	EDrosophila DNA for transposable element D near 3'e
PFASERAA	EP.falciparum serine-repeat antigen protein gene (S
DDIUBIRPB	ESlime mold (S.discoikeum) ubiquitin lambda-229 gen
DDIUDPGP	ED.discoideum UDP glucose pyrophosphorylase gene, c
NEMRDNAI	EAscaris lumbricoides rDNA with 185 rRNA gene 5'-en
PFAHRPC	EP.falciparum histidine-rich protein genes.
PFA412ANT	EP.falciparum 41-2 protein antigen, complete cds.
CHIRGNTS	EC.thummi piger non-transcribed spacer region of rR
PFACSP	EP.falciparum (Wellcome) circumsporozoite protein g
TRBVSG17B	Et.brucei viriant surface glycoprotein 117b mrna, S
DDIACT32	ESlime mold (D.oiscoldeum) actin 6 gene, 3' end.
CHIBR1G	EChironomus pallidivittatus BR1 gene for giant secr
DDIACT15P	EDictyostelium actin 15 gene promoter region.
PFAACTA	ED.falciparum pf-actin I gene encoding actin, compl
PFAHPRT	EPlasmodium mRNA for hypoxanthine-guanine phosphori

FIG. 8d

<u>GENBANK</u> <u>CODE</u>	<u>GENE</u>
DDIACT15	ED.discoideum actin 15 gene, complete cds.
PFAABRA	EPlasmodium falciparum p101/acidic basic repeat ant
TRCSLRCA	ET.cruzi small spliced leader (mini-exon) RNA gene
TRCSLRC	ETrypanosoma cruzi small spliced leader (mini-exon)
DDIACT24	ESlime mold (D.discoideum) actin 2-sub 2 pseudogene
BRPANTP	EB.malayi 63 kg antigen (potentially protective) ge
TRCKAP	ET.cruzi kinetoplast-associated protein (KAP) gene,
	EPlasmodium falciparum circumsporozoite (CS) protel

FIG. 8e

INTERNATIONAL SEARCH REPORT

International Application No. **PCT/US91/09402**

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all)³

According to International Patent Classification (IPC) or to both National Classification and IPC
 IPC (5): C12Q 1/70; C12Q 1/68; C12P 19/34; C07H 15/12; C07H 17/00
 US CL : 435/5, 6, 91, 810; 536/27, 28

II. FIELDS SEARCHED

Minimum Documentation Searched⁴

Classification System	Classification Symbols
U.S.	435/5, 6, 91, 810; 536/27, 28

Documentation Searched other than Minimum Documentation
to the extent that such Documents are included in the Fields Searched⁵

APS, CAS DATA BASES

III. DOCUMENTS CONSIDERED TO BE RELEVANT¹⁴

Category*	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁸
A	US, A, 4,683,195 (Mullis et al.) 28 July 1987.	1-62
Y	Science, volume 238, issued 30 October 1987, Moser et al., "Sequence-Specific Cleavage of Double Helical DNA by Triple-Helix Formation", pages 645-650, see abstract.	1-8, 11-43, 46-62
Y	Nucleic Acids Research, volume 16, No. 24, issued 1988; Francois et al., "Sequence Specific recognition of the major groove of DNA by oligodeoxynucleotides via triple helix formation. Footprinting studies," pages 11431-11440, see abstract.	1-8, 11-43, 46-62
Y	Proc. Nat. Acad. Sci., volume 85, issued June 1988; Kohui et al., "Magnesium ion-dependent triple-helix structure formed by homopurine-homopyrimidine sequences in supercoiled plasmid DNA" pages 3781-3786, see abstract.	1-9, 11-43, 46-62

* Special categories of cited documents:¹⁵

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search²
27 MARCH 1992

Date of Mailing of this International Search Report²
27 APR 1992

International Searching Authority¹
ISA/US

Signature of Authorized Officer²⁰
Scott A. Chambers
Scott A. Chambers

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